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(54) **NUCLEIC ACIDS ENCODING D4
DESATURASES AND D5 ELONGASES**

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13, 2007.

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C12N 9/02 (2006.01)

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CPC **C12N 9/1085** (2013.01); **A01K 67/027**
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9/1029 (2013.01)

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(57) **ABSTRACT**

Disclosed are methods and compositions related to ONC-
T18, D4-desaturases, D5 elongases, their isolation, charac-
terization, production, identification, and use for fatty acid
production, as well as organisms containing these compo-
sitions and organisms expressing them.

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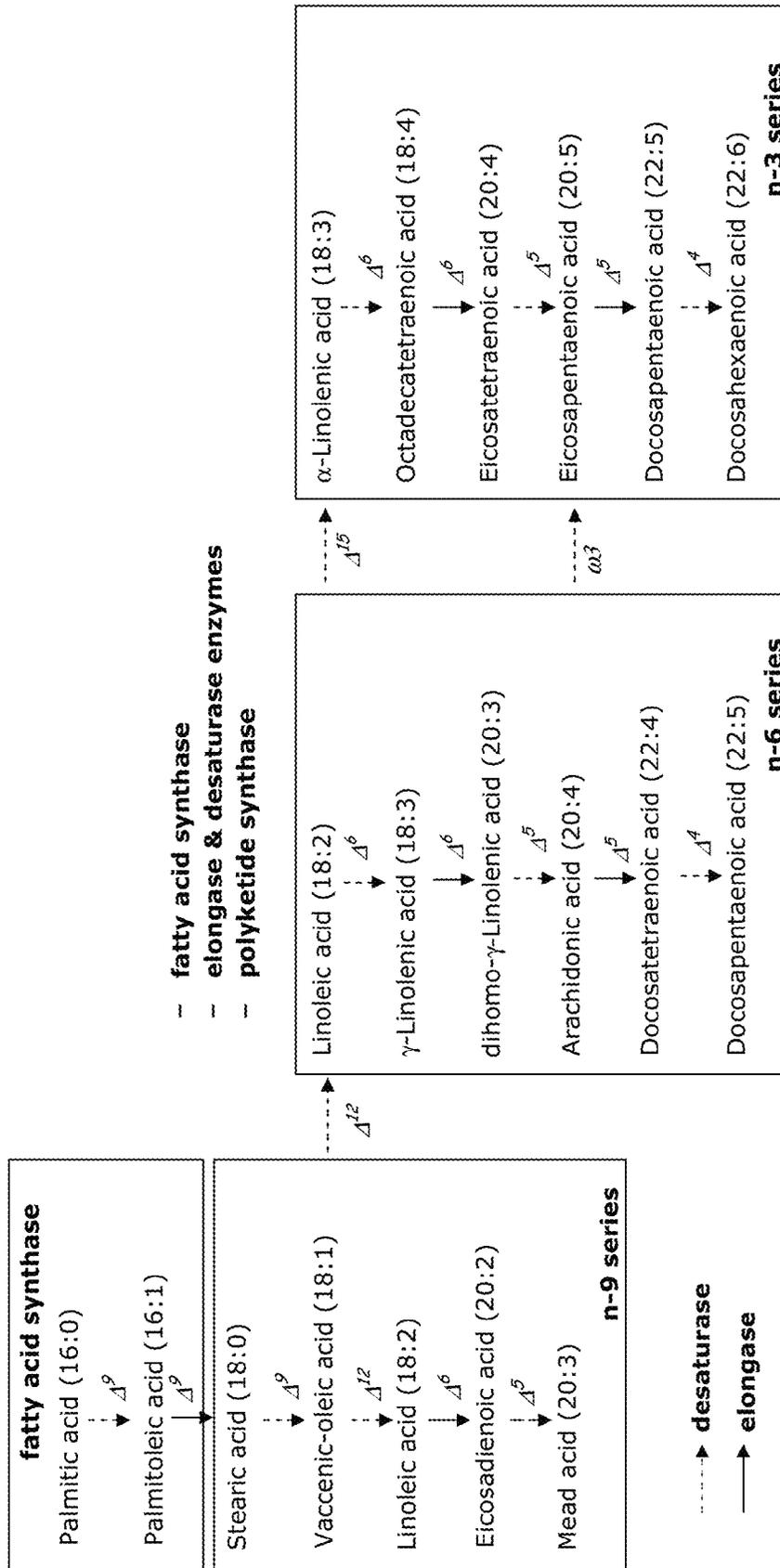


FIG. 1

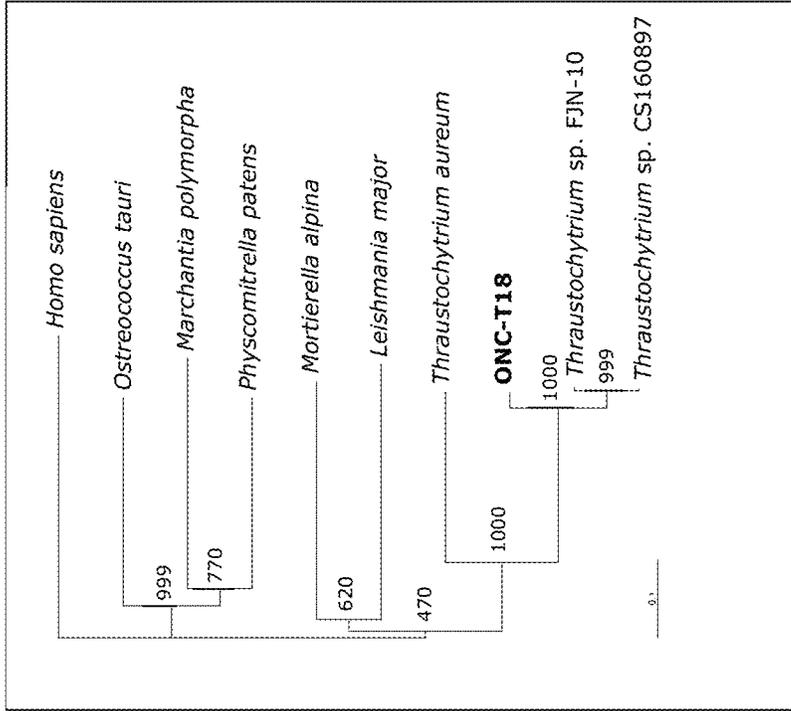


FIG. 2B

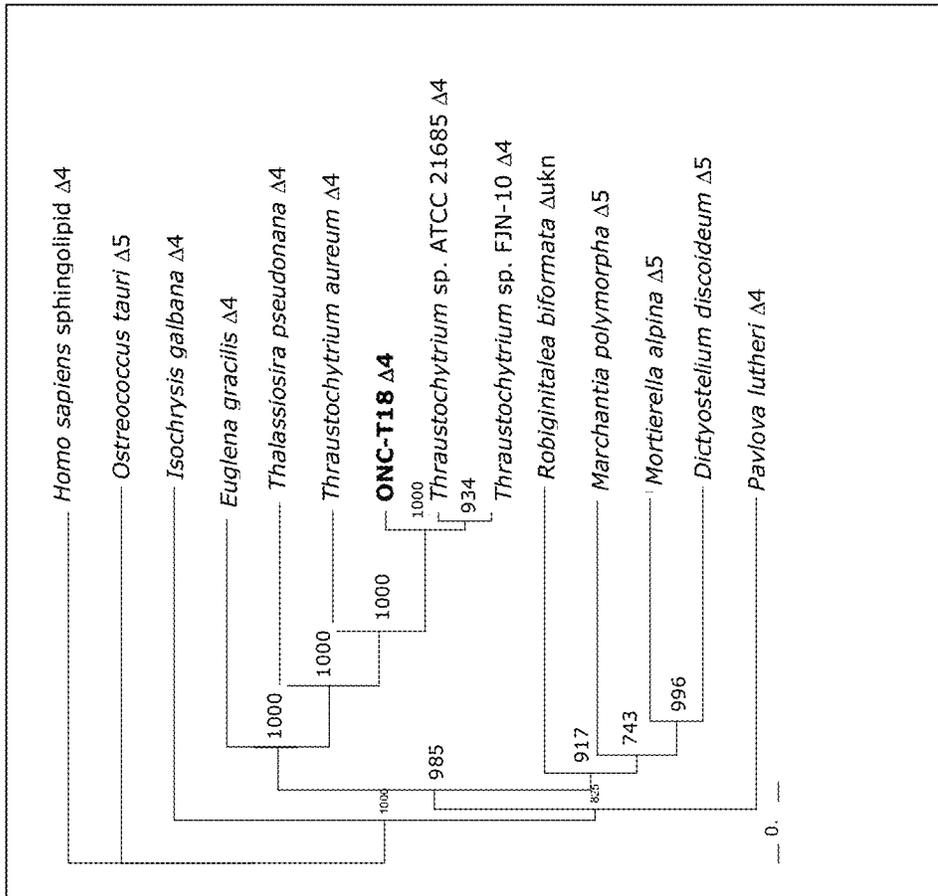


FIG. 2A

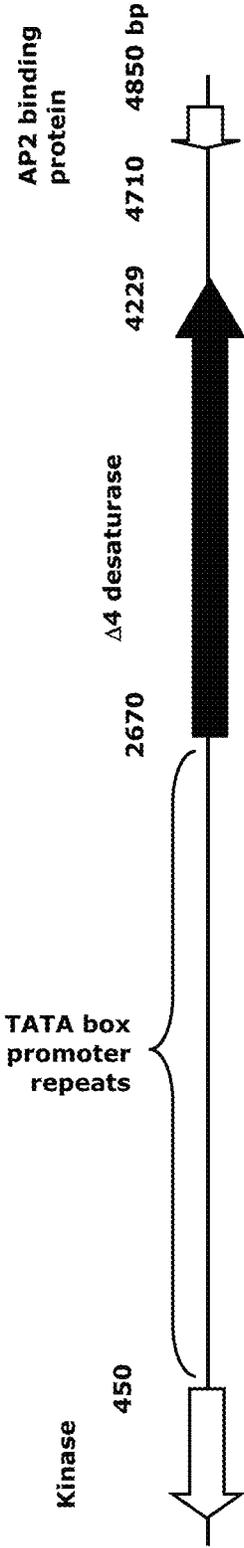


FIG. 3

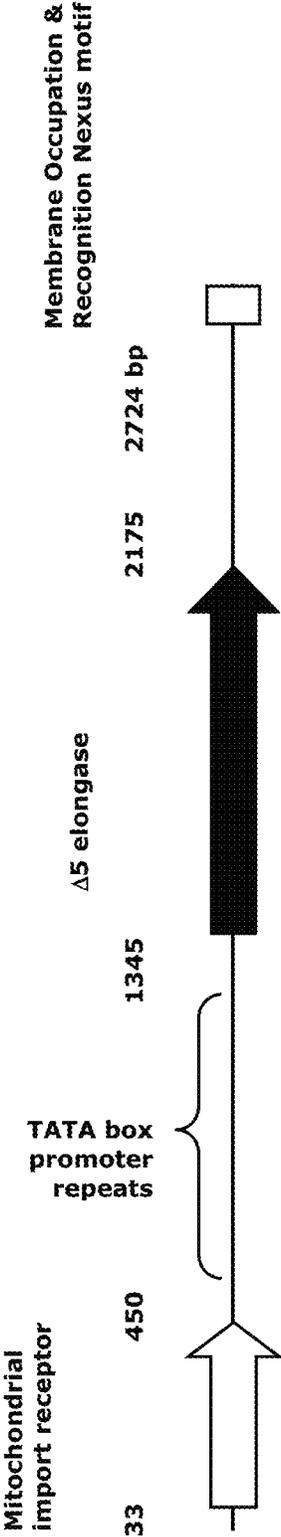


FIG. 4

NUCLEIC ACIDS ENCODING D4 DESATURASES AND D5 ELONGASES

CROSS REFERENCE TO RELATED APPLICATIONS

This application is a divisional application of U.S. patent application Ser. No. 12/668,793, filed Jun. 17, 2010, now U.S. Pat. No. 8,765,422, which is a national phase application claiming priority to International Patent Application No. PCT/IB2007/004553, filed Oct. 31, 2007, which claims the benefit of U.S. Provisional Patent Application No. 60/949,730, filed Jul. 13, 2007, each of which is incorporated herein by reference in its entirety.

I. BACKGROUND

There is overwhelming scientific evidence that (n-3) highly unsaturated fatty acids such as docosahexaenoic acid (DMA) have a positive effect on cardio-circulatory diseases, chronic inflammations and brain disorders. The (n-6) fatty acids such as eicosapentaenoic acid (EPA) on the other hand have been noted as intermediate metabolites within the eicosanoid steroids, such as prostaglandins, leucotrienes or the like.

Currently, the main source of these highly unsaturated fatty acids is fish, with EPA and DHA noted within various blue fish (such as sardines and tuna) at amounts around 20% and 10%, respectively. It is believed that such a fatty acid profile occurs through the natural selection of optimal ratios for optimal performance within each species of fish. Yet, if one intends to use fish oil as the sole source of these lipids, several disadvantages exist, such as problems with flavor taint, uncontrollable fluctuations in availability and natural fish oil content variability. In addition, if one intends to obtain a highly purified (n-3) or (n-6) oil from these sources, it is very difficult to preferentially separate and purify.

Previously disclosed is a Thraustochytriales eukaryote, ONC-T18 and related organisms, capable of producing high amounts of DHA and EPA as well as other preferred fatty acids. ONC-T18 is disclosed in International Application PCT/IB2006/003977 and United States provisional applications 60/751,401 and 60/821,084 which are all herein incorporated by reference for information at least related to ONC-T18 and fatty acids produced therein. The manipulation of the DHA and EPA pathways is desirable. Disclosed herein is the isolation and characterization of two enzymes from ONC-T18 involved in these pathways, a D4 desaturase and a D5 elongase.

II. SUMMARY

Disclosed are methods and compositions related to ONC-T18, D4-desaturases D5 elongases, their isolation, characterization, production, identification, and use for fatty acid production, as well as organisms containing these compositions and organisms expressing them.

III. BRIEF DESCRIPTION OF THE DRAWINGS

The accompanying drawings, which are incorporated in and constitute a part of this specification, illustrate several embodiments and together with the description illustrate the disclosed compositions and methods.

FIG. 1 shows the pathways of EPA and DHA production.

FIG. 2A shows a phylogentic tree for the isolated D4 desaturase and FIG. 2B shows a phlogenetic tree for the isolated D5-elongase.

FIG. 3 shows a genetic outline schematic for the isolated D4 desaturase.

FIG. 4 shows a genetic outline schematic for the isolated D5 elongase.

IV. DETAILED DESCRIPTION

Before the present compounds, compositions, articles, devices, and/or methods are disclosed and described, it is to be understood that they are not limited to specific synthetic methods or specific recombinant biotechnology methods unless otherwise specified, or to particular reagents unless otherwise specified, as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting.

A. Definitions

As used in the specification and the appended claims, the singular forms “a,” “an” and “the” include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to “a pharmaceutical carrier” includes mixtures of two or more such carriers, and the like.

Ranges can be expressed herein as from “about” one particular value, and/or to “about” another particular value. When such a range is expressed, another embodiment includes from the one particular value and/or to the other particular value. Similarly, when values are expressed as approximations, by use of the antecedent “about,” it will be understood that the particular value forms another embodiment. It will be further understood that the endpoints of each of the ranges are significant both in relation to the other endpoint, and independently of the other endpoint. It is also understood that there are a number of values disclosed herein, and that each value is also herein disclosed as “about” that particular value in addition to the value itself. For example, if the value “10” is disclosed, then “about 10” is also disclosed. It is also understood that when a value is disclosed that “less than or equal to” the value, “greater than or equal to the value” and possible ranges between values are also disclosed, as appropriately understood by the skilled artisan. For example, if the value “10” is disclosed the “less than or equal to 10” as well as “greater than or equal to 10” is also disclosed. It is also understood that the throughout the application, data is provided in a number of different formats, and that this data, represents endpoints and starting points, and ranges for any combination of the data points. For example, if a particular data point “10” and a particular data point 15 are disclosed, it is understood that greater than, greater than or equal to, less than, less than or equal to, and equal to 10 and 15 are considered disclosed as well as between 10 and 15. It is also understood that each unit between two particular units are also disclosed. For example, if 10 and 15 are disclosed, then 11, 12, 13, and 14 are also disclosed.

In this specification and in the claims which follow, reference will be made to a number of terms which shall be defined to have the following meanings:

“Optional” or “optionally” means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances where it does not.

“Primers” are a subset of probes which are capable of supporting some type of enzymatic manipulation and which

can hybridize with a target nucleic acid such that the enzymatic manipulation can occur. A primer can be made from any combination of nucleotides or nucleotide derivatives or analogs available in the art which do not interfere with the enzymatic manipulation.

"Probes" are molecules capable of interacting with a target nucleic acid, typically in a sequence specific manner, for example through hybridization. The hybridization of nucleic acids is well understood in the art and discussed herein. Typically a probe can be made from any combination of nucleotides or nucleotide derivatives or analogs available in the art.

Throughout this application, various publications are referenced. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this pertains. The references disclosed are also individually and specifically incorporated by reference herein for the material contained in them that is discussed in the sentence in which the reference is relied upon.

Disclosed are the components to be used to prepare the disclosed compositions as well as the compositions themselves to be used within the methods disclosed herein. These and other materials are disclosed herein, and it is understood that when combinations, subsets, interactions, groups, etc. of these materials are disclosed that while specific reference of each various individual and collective combinations and permutation of these compounds may not be explicitly disclosed, each is specifically contemplated and described herein. For example, if a particular D4 desaturase and D5 elongase is disclosed and discussed and a number of modifications that can be made to a number of molecules including the D4 desaturase and D5 elongase are discussed, specifically contemplated is each and every combination and permutation of D4 desaturase and D5 elongase and the modifications that are possible unless specifically indicated to the contrary. Thus, if a class of molecules A, B, and C are disclosed as well as a class of molecules D, E, and F and an example of a combination molecule, A-D is disclosed, then even if each is not individually recited each is individually and collectively contemplated meaning combinations, A-E, A-F, B-D, B-E, B-F, C-D, C-E, and C-F are considered disclosed. Likewise, any subset or combination of these is also disclosed. Thus, for example, the sub-group a A-E, B-F, and C-E would be considered disclosed. This concept applies to all aspects of this application including, but not limited to, steps in methods of making and using the disclosed compositions. Thus, if there are a variety of additional steps that can be performed it is understood that each of these additional steps can be performed with any specific embodiment or combination of embodiments of the disclosed methods.

B. Compositions

The use of omega-3 concentrates (eicosapentaenoic acid EPA, 20:5 n-3 and docosahexaenoic acid DHA, 22:6 n-3) has become important in the fortification of certain foods to promote a healthy diet. *Thraustochytrids* are marine protists that naturally produce DHA, at up to 20% of their biomass. The capability to ferment these organisms provides for a renewable, long-term source of these omega-3 oils. Characterisation of the fatty acid metabolic pathway (FIG. 1) reveals the importance of a D5-elongase responsible for the elongation of EPA into docosapentaenoic acid (DPA, 22:5 n-3), as well as a D4-desaturase being involved in the desaturation of DPA into DHA. Manipulation of the specific enzyme activity can influence the yield of EPA and DHA

produced by our ONC-T18 strain as required by the needs of the market, and our customers.

Using degenerate primers constructed from a conserved region in the gene sequence from enzymes of different *Thraustochytrid* strains (*Thraustochytrium* sp. (CS020087), *Thraustochytrium aureum* (AF391546), *Thraustochytrium* sp. ATCC 34304 (AF391543), *Thraustochytrium* sp. ATCC 21685 (AF489589), and *Thraustochytrium* sp. FJN-10 (DQ133575) for D4-desaturase; *Thraustochytrium* sp. (CS160897) and *Thraustochytrium aureum* (CS160879) for D5-elongase) a D4-desaturase and a D5 elongase from ONC T-18 were isolated. Further a portion of the genes from genomic DNA (967 bp and 593 bp, respectively) was PCR amplified. Genome walking (APAgene GOLD kit, BIO S&T, Montreal, Quebec) was used to extend the known sequence to incorporate the entire open reading frame, and extend further to identify the adjacent genes on either side along with the promoter region. Primers were then constructed for the complete sequence to produce a full gene PCR product incorporating the entire open reading frame (1758 bp and 1099 bp, respectively). The PCR product was cloned into pT7-Blue3 vector (Novagen, San Diego, Calif.) and transformed into *Escherichia coli* NovaBlue (DE3) (Novagen, San Diego, Calif.) and then sequenced.

Resultant gene plasmids were purified using the Ultra-Clean 6 Minute Mini Plasmid Prep kit (MC) BIO Laboratories, Inc., Solana Beach, Calif.). The gene inserts were then excised using the restriction enzymes BamHI and NotI and cloned into the pYES2 yeast expression vector (Invitrogen, Carlsbad, Calif.). The new vector constructs identified as pYDes (D4-desaturase) and pYElo (D5-elongase) were then transformed into *Saccharomyces cerevisiae* INVSc1 using the S.c. EasyComp Transformation kit (Invitrogen, Carlsbad, Calif.), under the galactose promoter Gal1. The negative control strain was INVSc1 containing the unaltered pYES2 vector, and these were grown simultaneously. The vector selection was done using the uracil auxotrophy of the yeast strain, SC medium without uracil was used.

The activity and specificity of the D4-desaturase and D5 elongase were determined using the yeast expression system. The transformed yeast was grown in SC-U medium containing 2% glucose, 1% Tergitol NP-40 for 48 hrs, 150 RPM at 30° C. A substrate medium was prepared containing SC-U, 2% galactose, 1% raffinose, 1% Tergitol NP-40 and 500 μM specific free fatty acid. The transformed yeast culture was inoculated into 100 ml substrate medium at an OD600 of 0.5 and cultures incubated at 20° C. for 5 days. The biomass was recovered by centrifugation at 2000 RPM for 5 min, washed once with 100 mM phosphate buffer (pH 7.0), then freeze dried. Fatty acid methyl ester gas chromatography was subsequently carried out to determine the efficiency of fatty acid desaturation and elongation. The percent conversion of the substrate was determined by calculating (product)/(substrate+product)*100.

BLASTx results showed the ONC T-18 D4 desaturase to be 96% similar to a *Thraustochytrium* sp. ATCC 21685 D4-desaturase. FIG. 2A shows a rooted neighbour-joining phylogenetic tree, determined using ClustalX, bootstrap analysis (1000x) using the results of a BLASTx search when compared to the ONC-T18 D4-desaturase sequence. The D4-desaturase gene has an open reading frame of 1560 bp, transcribing a 519 amino acid protein. Analysis of this protein shows a cytochrome b5 domain with three histidine box motifs and four transmembrane regions, all elements characteristic of front-end desaturase. Adjacent to this gene are five putative TATA boxes, multiple repeat regions, two

promoters and a protein kinase identified upstream and an AP2 binding protein downstream (FIG. 3).

Conversely, a BLASTx search identified the D5-elongase protein as having 89% identity to a *Thraustochytrium* sp. FJN-10 polyunsaturated fatty acid elongase. FIG. 2B shows a rooted neighbour-joining phylogenetic tree, determined using ClustalX, bootstrap analysis (1000×) using the results of a BLASTx search when compared to the ONC-T18 D5-elongase sequence. Further analysis determined that this 831 bp long elongase, coding a 276 amino acid protein, contains four transmembrane regions specific to mitochondria, and one histidine box motif. The upstream component of this D5-elongase region comprises a single TATA box, multiple repeat regions and a promoter prior to a mitochondrial import receptor, while downstream the beginning of a membrane occupation and recognition nexus motif was identified (FIG. 4).

Characterisation of pYDes (Table 1) for both the n-3 and n-6 pathways, showed a 14% conversion of DPA n-3 or docosapentaenoic acid (DTA 22:4 n-6) to DIM or DPA n-6, respectively.

TABLE 1

D4-desaturase enzyme activity and characterisation						
Substrate	Product	% conversion			With 0.01% ferric citrate	
		average	stdev	(x)	% conversion	increase
DPA	DHA	14.04	4.01	4	38.70	2.75 fold
DTA	DPA n-6	13.76	1.31	3	33.43	2.43 fold
DGLA	ARA	0.87	0.27	3		

When fed the corresponding D4-desaturase substrate dihomogamma-linolenic acid (DGLA, 20:3 n-6), no conversion was detected. In an effort to increase pYDes activity, trace metals such as ferric citrate was added to the media, resulting in an increase in DPA to DHA conversion (Table 1). In contrast, presently pYelo shows minimal conversion when either EPA or arachidonic acid (ARA 20:4 n-6) were fed.

D4-desaturase and D5-elongase genes from the high fatty acid producing strain *Thraustochytrium* sp. ONC-T18, have been successfully isolated and cloned, followed by expression in *S. cerevisiae*.

Furthermore, the D4-desaturase enzyme was shown to convert both DPA or DTA to their respective end products both in their native form and via supplementation with trace metals. Feed studies with other fatty acids confirmed the D4 specific activity of this desaturase. Through the use of gene manipulation techniques, such as error-prone PCR, this activity will be further enhanced so as to effect an increase in production of DHA in our strain.

Disclosed are compositions comprising a D4 desaturase wherein the D4 desaturase has at least or greater than 70%, 80%, 89%, 90%, 95%, 96%, 97% identity to SEQ ID NO:26.

Also disclosed are compositions, wherein any change away from SEQ ID NO:26 is a conservative change.

Also disclosed are compositions comprising a nucleic acid wherein the nucleic acid encodes any of the D4 desaturase.

Also disclosed are compositions, further comprising a vector.

Also disclosed are compositions comprising a cell wherein the cell comprises any of the compositions.

Also disclosed are compositions, wherein the cell is a eukaryote, a prokaryote, a *Thraustochytrid*, a yeast, or an *E. coli*.

Also disclosed are compositions comprising a non-human animal wherein the non-human animal comprises any of the compositions.

Also disclosed are compositions, wherein the composition produces more polyunsaturated fatty acids than the composition in the absence of the D4 desaturase.

Also disclosed are compositions, wherein the fatty acid is EPA or DHA.

Also disclosed are compositions, wherein the desaturase contains at least one histidine box.

Also disclosed are compositions, wherein the desaturase contains at least 2 histidine boxes.

Also disclosed are compositions, wherein the desaturase contains at least three histidine boxes.

Also disclosed are compositions, wherein the histidine box comprises the sequence HXXHH (SEQ ID NO:29) where X is any amino acid.

Also disclosed are compositions, wherein the histidine box comprises the sequence QXXHH (SEQ ID NO:30).

Also disclosed are compositions, wherein the desaturase also comprises a cytochrome b5 domain.

Also disclosed are compositions, wherein the cytochrome b5 domain resides at the 5'-end.

Also disclosed are compositions, wherein the desaturase is in the presence of a desaturase substrate

Also disclosed are compositions, wherein the substrate has a concentration of at least 100 μM, 200 μM, 300 μM, 400 μM, 500 μM, 600 μM, 700 μM, 800 μM, 900 μM, or 1000 μM.

Also disclosed are compositions, wherein the substrate is Docosapentaenoic acid (22:5n-3), Docosatetraenoic acid (22:4n-6), or Dihomo-gamma-linolenic acid (20:3n-6).

Also disclosed are compositions, wherein the desaturase converts at least 0.1%, 0.5%, 1%, 5%, 10%, 30%, 50%, 70%, 90%, 95% of the available substrate.

Also disclosed are compositions, wherein the desaturase converts the amount of substrate shown in Table 1.

Also disclosed are compositions, wherein the composition is isolated.

Also disclosed are compositions comprising a D5 elongase wherein the D5 elongase has at least or greater than 70%, 80%, 89%, 90%, 95%, 96%, 97% identity to SEQ ID NO:15.

Also disclosed are compositions, wherein any change away from SEQ ID NO: 15 is a conservative change.

Also disclosed are compositions comprising a nucleic acid wherein the nucleic acid encodes any of the D5 elongases.

Also disclosed are compositions encoding elongases or desaturases, further comprising a vector.

Also disclosed are compositions wherein the composition produces more polyunsaturated fatty acids than the composition in the absence of the D5 elongase, such as DHA or EPA.

Also disclosed are compositions wherein the elongase is in the presence of an elongase substrate

Also disclosed are compositions, wherein the substrate is Eicosapentaenoic acid (20:5n-3) or Arachidonic acid (20:4n-6).

Also disclosed are compositions, wherein the elongase converts at least 0.1%, 0.5%, 1%, 5%, 10%, 30%, 50%, 70%, 90%, 95% of the available substrate.

Also disclosed are compositions, wherein the composition is isolated.

Also disclosed are compositions comprising any of the disclosed desaturase compositions and any of the disclosed elongase compositions.

Also disclosed are methods for producing a polyunsaturated fatty acid comprising using one or more of any of the compositions.

Also disclosed are methods, wherein the fatty acid produced is either EPA or DHA.

Also disclosed are methods of producing the compositions comprising isolating any of the compositions.

1. Sequence Similarities

It is understood that as discussed herein the use of the terms homology and identity mean the same thing as similarity. Thus, for example, if the use of the word homology is used between two non-natural sequences it is understood that this is not necessarily indicating an evolutionary relationship between these two sequences, but rather is looking at the similarity or relatedness between their nucleic acid sequences. Many of the methods for determining homology between two evolutionarily related molecules are routinely applied to any two or more nucleic acids or proteins for the purpose of measuring sequence similarity regardless of whether they are evolutionarily related or not.

In general, it is understood that one way to define any known variants and derivatives or those that might arise, of the disclosed genes and proteins herein, is through defining the variants and derivatives in terms of homology to specific known sequences. This identity of particular sequences disclosed herein is also discussed elsewhere herein. In general, variants of genes and proteins herein disclosed typically have at least, about 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, or 99 percent homology to the stated sequence or the native sequence. Those of skill in the art readily understand how to determine the homology of two proteins or nucleic acids, such as genes. For example, the homology can be calculated after aligning the two sequences so that the homology is at its highest level.

Another way of calculating homology can be performed by published algorithms. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, PASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by inspection.

The same types of homology can be obtained for nucleic acids by for example the algorithms disclosed in Zuker, *M. Science* 244:48-52, 1989, Jaeger et al. *Proc. Natl. Acad. Sci. USA* 86:7706-7710, 1989, Jaeger et al. *Methods Enzymol.* 183:281-306, 1989 which are herein incorporated by reference for at least material related to nucleic acid alignment. It is understood that any of the methods typically can be used and that in certain instances the results of these various methods may differ, but the skilled artisan understands if identity is found with at least one of these methods, the sequences would be said to have the stated identity, and be disclosed herein.

For example, as used herein, a sequence recited as having a particular percent homology to another sequence refers to sequences that have the recited homology as calculated by any one or more of the calculation methods described above. For example, a first sequence has 80 percent homology, as defined herein, to a second sequence if the first sequence is calculated to have 80 percent homology to the second sequence using the Zuker calculation method even if the first

sequence does not have 80 percent homology to the second sequence as calculated by any of the other calculation methods. As another example, a first sequence has 80 percent homology, as defined herein, to a second sequence if the first sequence is calculated to have 80 percent homology to the second sequence using both the Zuker calculation method and the Pearson and Lipman calculation method even if the first sequence does not have 80 percent homology to the second sequence as calculated by the Smith and Waterman calculation method, the Needleman and Wunsch calculation method, the Jaeger calculation methods, or any of the other calculation methods. As yet another example, a first sequence has 80 percent homology, as defined herein, to a second sequence if the first sequence is calculated to have 80 percent homology to the second sequence using each of calculation methods (although, in practice, the different calculation methods will often result in different calculated homology percentages).

2. Hybridization/Selective Hybridization

The term hybridization typically means a sequence driven interaction between at least two nucleic acid molecules, such as a primer or a probe and a gene. Sequence driven interaction means an interaction that occurs between two nucleotides or nucleotide analogs or nucleotide derivatives in a nucleotide specific manner. For example, G interacting with C or A interacting with T are sequence driven interactions. Typically sequence driven interactions occur on the Watson-Crick face or Hoogsteen face of the nucleotide. The hybridization of two nucleic acids is affected by a number of conditions and parameters known to those of skill in the art. For example, the salt concentrations, pH, and temperature of the reaction all affect whether two nucleic acid molecules will hybridize.

Parameters for selective hybridization between two nucleic acid molecules are well known to those of skill in the art. For example, in some embodiments selective hybridization conditions can be defined as stringent hybridization conditions. For example, stringency of hybridization is controlled by both temperature and salt concentration of either or both of the hybridization and washing steps. For example, the conditions of hybridization to achieve selective hybridization may involve hybridization in high ionic strength solution (6×SSC or 6×SSPE) at a temperature that is about 12-25° C. below the T_m (the melting temperature at which half of the molecules dissociate from their hybridization partners) followed by washing at a combination of temperature and salt concentration chosen so that the washing temperature is about 5° C. to 20° C. below the T_m. The temperature and salt conditions are readily determined empirically in preliminary experiments in which samples of reference DNA immobilized on filters are hybridized to a labeled nucleic acid of interest and then washed under conditions of different stringencies. Hybridization temperatures are typically higher for DNA-RNA and RNA-RNA hybridizations. The conditions can be used as described above to achieve stringency, or as is known in the art. (Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1989; Kunkel et al. *Methods Enzymol.* 1987:154: 367, 1987 which is herein incorporated by reference for material at least related to hybridization of nucleic acids). A preferable stringent hybridization condition for a DNA: DNA hybridization can be at about 68° C. (in aqueous solution) in 6×SSC or 6×SSPE followed by washing at 68° C. Stringency of hybridization and washing, if desired, can be reduced accordingly as the degree of complementarity desired is decreased, and further, depending upon the G-C or

A-T richness of any area wherein variability is searched for. Likewise, stringency of hybridization and washing, if desired, can be increased accordingly as homology desired is increased, and further, ID depending upon the G-C or A-T richness of any area wherein high homology is desired, all as known in the art.

Another way to define selective hybridization is by looking at the amount (percentage) of one of the nucleic acids bound to the other nucleic acid. For example, in some embodiments selective hybridization conditions would be when at least about, 60, 65, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of the limiting nucleic acid is bound to the non-limiting nucleic acid. Typically, the non-limiting primer is in for example, 10 or 100 or 1000 fold excess. This type of assay can be performed at under conditions where both the limiting and non-limiting primer are for example, 10 fold or 100 fold or 1000 fold below their k_d , or where only one of the nucleic acid molecules is 10 fold or 100 fold or 1000 fold or where one or both nucleic acid molecules are above their k_d .

Another way to define selective hybridization is by looking at the percentage of primer that gets enzymatically manipulated under conditions where hybridization is required to promote the desired enzymatic manipulation. For example, in some embodiments selective hybridization conditions would be when at least about, 60, 65, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of the primer is enzymatically manipulated under conditions which promote the enzymatic manipulation, for example if the enzymatic manipulation is DNA extension, then selective hybridization conditions would be when at least about 60, 65, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100 percent of the primer molecules are extended. Preferred conditions also include those suggested by the manufacturer or indicated in the art as being appropriate for the enzyme performing the manipulation.

Just as with homology, it is understood that there are a variety of methods herein disclosed for determining the level of hybridization between two nucleic acid molecules. It is understood that these methods and conditions may provide different percentages of hybridization between two nucleic acid molecules, but unless otherwise indicated meeting the parameters of any of the methods would be sufficient. For example if 80% hybridization was required and as long as hybridization occurs within the required parameters in any one of these methods it is considered disclosed herein.

It is understood that those of skill in the art understand that if a composition or method meets any one of these criteria for determining hybridization either collectively or singly it is a composition or method that is disclosed herein.

3. Nucleic Acids

There are a variety of molecules disclosed herein that are nucleic acid based, including for example the nucleic acids that encode, for example, the isolated D4 desaturase and D5 elongase as well as any other proteins disclosed herein, as well as various functional nucleic acids. The disclosed nucleic acids are made up of for example, nucleotides, nucleotide analogs, or nucleotide substitutes. Non-limiting examples of these and other molecules are discussed herein. It is understood that for example, when a vector is expressed in a cell, that the expressed mRNA will typically be made up of A, C, G, and U. Likewise, it is understood that it for example, an antisense molecule is introduced into a cell or cell environment through for example exogenous delivery, it

is advantageous that the antisense molecule be made up of nucleotide analogs that reduce the degradation of the antisense molecule in the cellular environment.

a) Nucleotides and Related Molecules

A nucleotide is a molecule that contains a base moiety, a sugar moiety and a phosphate moiety. Nucleotides can be linked together through their phosphate moieties and sugar moieties creating an internucleoside linkage. The base moiety of a nucleotide can be adenin-9-yl (A), cytosin-1-yl (C), guanin-9-yl (G), uracil-1-yl (U), and thymine-1-yl (T). The sugar moiety of a nucleotide is a ribose or a deoxyribose. The phosphate moiety of a nucleotide is pentavalent phosphate. A non-limiting example of a nucleotide would be 3'-AMP (3'-adenosine monophosphate) or 5'-GMP (5'-guanosine monophosphate).

A nucleotide analog is a nucleotide which contains some type of modification to either the base, sugar, or phosphate moieties. Modifications to nucleotides are well known in the art and would include for example, 5-methylcytosine (5-me-C), 5-hydroxymethyl cytosine, xanthine, hypoxanthine, and 2-aminoadenine as well as modifications at the sugar or phosphate moieties.

Nucleotide substitutes are molecules having similar functional properties to nucleotides, but which do not contain a phosphate moiety, such as peptide nucleic acid (PNA). Nucleotide substitutes are molecules that will recognize nucleic acids in a Watson-Crick or Hoogsteen manner, but which are linked together through a moiety other than a phosphate moiety. Nucleotide substitutes are able to conform to a double helix type structure when interacting with the appropriate target nucleic acid.

It is also possible to link other types of molecules (conjugates) to nucleotides or nucleotide analogs to enhance for example, cellular uptake. Conjugates can be chemically linked to the nucleotide or nucleotide analogs. Such conjugates include but are not limited to lipid moieties such as a cholesterol moiety. Letsinger et al., Proc. Natl. Acad. Sci. USA, 1989, 86, 6553-6556).

A Watson-Crick interaction is at least one interaction with the Watson-Crick face of a nucleotide, nucleotide analog, or nucleotide substitute. The Watson-Crick face of a nucleotide, nucleotide analog, or nucleotide substitute includes the C2, N1, and C6 positions of a purine based nucleotide, nucleotide analog, or nucleotide substitute and the C2, N3, C4 positions of a pyrimidine based nucleotide, nucleotide analog, or nucleotide substitute.

A Hoogsteen interaction is the interaction that takes place on the Hoogsteen face of a nucleotide or nucleotide analog, which is exposed in the major groove of duplex DNA. The Hoogsteen face includes the N7 position and reactive groups (NH2 or O) at the C6 position of purine nucleotides.

b) Sequences

There are a variety of sequences related to, for example, the isolated D4 desaturase and D5 elongase as well as any other protein disclosed herein that are disclosed on Genbank, and these sequences and others are herein incorporated by reference in their entireties as well as for individual subsequences contained therein.

A variety of sequences are provided herein and these and others can be found in Genbank, at www.pubmed.gov. Those of Skill in the art understand how to resolve sequence discrepancies and differences and to adjust the compositions and methods relating to a particular sequence to other related sequences. Primers and/or probes can be designed for any sequence given the information disclosed herein and known in the art.

c) Primers and Probes

Disclosed are compositions including primers and probes, which are capable of interacting with the genes disclosed herein. In certain embodiments the primers are used to support DNA amplification reactions. Typically the primers will be capable of being extended in a sequence specific manner. Extension of a primer in a sequence specific manner includes any methods wherein the sequence and/or composition of the nucleic acid molecule to which the primer is hybridized or otherwise associated directs or influences the composition or sequence of the product produced by the extension of the primer. Extension of the primer in a sequence specific manner therefore includes, but is not limited to, PCR, DNA sequencing, DNA extension, DNA polymerization, RNA transcription, or reverse transcription. Techniques and conditions that amplify the primer in a sequence specific manner are preferred. In certain embodiments the primers are used for the DNA amplification reactions, such as PCR or direct sequencing. It is understood that in certain embodiments the primers can also be extended using non-enzymatic techniques, where for example, the nucleotides or oligonucleotides used to extend the primer are modified such that they will chemically react to extend the primer in a sequence specific manner. Typically the disclosed primers hybridize with the nucleic acid or region of the nucleic acid or they hybridize with the complement of the nucleic acid or complement of a region of the nucleic acid.

d) Functional Nucleic Acids.

Functional nucleic acids are nucleic acid molecules that have a specific function, such as binding a target molecule or catalyzing a specific reaction. Functional nucleic acid molecules can be divided into the following categories, which are not meant to be limiting. For example, functional nucleic acids include antisense molecules, aptamers, ribozymes, triplex forming molecules, and external guide sequences. The functional nucleic acid molecules can act as effectors, inhibitors, modulators, and stimulators of a specific activity possessed by a target molecule, or the functional nucleic acid molecules can possess a de novo activity independent of any other molecules.

Functional nucleic acid molecules can interact with any macromolecule, such as DNA, RNA, polypeptides, or carbohydrate chains. Thus, functional nucleic acids can interact with the mRNA of the isolated D4 desaturase and D5 elongase or the genomic DNA of the isolated D4 desaturase and D5 elongase or they can interact with the polypeptide or fragments of the isolated D4 desaturase and D5 elongase. Often functional nucleic acids are designed to interact with other nucleic acids based on sequence homology between the target molecule and the functional nucleic acid molecule. In other situations, the specific recognition between the functional nucleic acid molecule and the target molecule is not based on sequence homology between the functional nucleic acid molecule and the target molecule, but rather is based on the formation of tertiary structure that allows specific recognition to take place.

Antisense molecules are designed to interact with a target nucleic acid molecule through either canonical or non-canonical base pairing. The interaction of the antisense molecule and the target molecule is designed to promote the destruction of the target molecule through, for example, RNaseH mediated RNA-DNA hybrid degradation. Alternatively the antisense molecule is designed to interrupt a processing function that normally would take place on the target molecule, such as transcription or replication. Antisense molecules can be designed based on the sequence of

the target molecule. Numerous methods for optimization of antisense efficiency by finding the most accessible regions of the target molecule exist. Exemplary methods would be in vitro selection experiments and DNA modification studies using DMS and DEPC. It is preferred that antisense molecules bind the target molecule with a dissociation constant (k_d) less than or equal to 10^{-6} , 10^{-8} , 10^{-10} or 10^{-12} . A representative sample of methods and techniques which aid in the design and use of antisense molecules can be found in the following non-limiting list of U.S. Pat. Nos. 5,135,917, 5,294,533, 5,627,158, 5,641,754, 5,691,317, 5,780,607, 5,786,138, 5,849,903, 5,856,103, 5,919,772, 5,955,590, 5,990,088, 5,994,320, 5,998,602, 6,005,095, 6,007,995, 6,013,522, 6,017,898, 6,018,042, 6,025,198, 6,033,910, 6,040,296, 6,046,004, 6,046,319, and 6,057,437.

Aptamers are molecules that interact with a target molecule, preferably in a specific way. Typically aptamers are small nucleic acids ranging from 15-50 bases in length that fold into defined secondary and tertiary structures, such as stem-loops or G-quartets. Aptamers can bind small molecules, such as ATP (U.S. Pat. No. 5,631,146) and theophylline (U.S. Pat. No. 5,580,737), as well as large molecules, such as reverse transcriptase (U.S. Pat. No. 5,786,462) and thrombin (U.S. Pat. No. 5,543,293). Aptamers can bind very tightly with k_d s from the target molecule of less than 10^{-12} M. It is preferred that the aptamers bind the target molecule with a k_d less than 10^{-6} , 10^{-8} , 10^{-10} , or 10^{-12} . Aptamers can bind the target molecule with a very high degree of specificity. For example, aptamers have been isolated that have greater than a 10000 fold difference in binding affinities between the target molecule and another molecule that differ at only a single position on the molecule (U.S. Pat. No. 5,543,293). It is preferred that the aptamer have a k_d with the target molecule at least 10, 100, 1000, 10,000, or 100,000 fold lower than the k_d with a background binding molecule. It is preferred when doing the comparison for a polypeptide for example, that the background molecule be a different polypeptide. For example, when determining the specificity of the isolated D4 desaturase and D5 elongase aptamers, the background protein could be serum albumin. Representative examples of how to make and use aptamers to bind a variety of different target molecules can be found in the following non-limiting list of U.S. Pat. Nos. 5,476,766, 5,503,978, 5,631,146, 5,731,424, 5,780,228, 5,792,613, 5,795,721, 5,846,713, 5,858,660, 5,861,254, 5,864,026, 5,869,641, 5,958,691, 6,001,988, 6,011,020, 6,013,443, 6,020,130, 6,028,186, 6,030,776, and 6,051,698.

Ribozymes are nucleic acid molecules that are capable of catalyzing a chemical reaction, either intramolecularly or intermolecularly. Ribozymes are thus catalytic nucleic acid. It is preferred that the ribozymes catalyze intermolecular reactions. There are a number of different types of ribozymes that catalyze nuclease or nucleic acid polymerase type reactions which are based on ribozymes found in natural systems, such as hammerhead ribozymes, (for example, but not limited to the following U.S. Pat. Nos. 5,334,711, 5,436,330, 5,616,466, 5,633,133, 5,646,020, 5,652,094, 5,712,384, 5,770,715, 5,856,463, 5,861,288, 5,891,683, 5,891,684, 5,985,621, 5,989,908, 5,998,193, 5,998,203, WO 9858058 by Ludwig and Sproat, WO 9858057 by Ludwig and Sproat, and WO 9718312 by Ludwig and Sproat) hairpin ribozymes (for example, but not limited to the following U.S. Pat. Nos. 5,631,115, 5,646,031, 5,683,902, 5,712,384, 5,856,188, 5,866,701, 5,869,339, and 6,022,962), and tetrahymena ribozymes (for example, but not limited to the following U.S. Pat. Nos. 5,595,873 and 5,652,107). There are also a number of ribozymes that are

not found in natural systems, but which have been engineered to catalyze specific reactions de novo (for example, but not limited to the following U.S. Pat. Nos. 5,580,967, 5,688,670, 5,807,718, and 5,910,408). Preferred ribozymes cleave RNA or DNA substrates, and more preferably cleave RNA substrates. Ribozymes typically cleave nucleic acid substrates through recognition and binding of the target substrate with subsequent cleavage. This recognition is often based mostly on canonical or non-canonical base pair interactions. This property makes ribozymes particularly good candidates for target specific cleavage of nucleic acids because recognition of the target substrate is based on the target substrates sequence. Representative examples of how to make and use ribozymes catalyze a variety of different reactions can be found in the following non-limiting list of U.S. Pat. Nos. 5,646,042, 5,693,535, 5,731,295, 5,811,300, 5,837,855, 5,869,253, 5,877,021, 5,877,022, 5,972,699, 5,972,704, 5,989,906, and 6,017,756.

Triplex forming functional nucleic acid molecules are molecules that can interact with either double-stranded or single-stranded nucleic acid. When triplex molecules interact with a target region, a structure called a triplex is formed, in which there are three strands of DNA forming a complex dependant on both Watson-Crick and Hoogsteen base-pairing. Triplex molecules are preferred because they can bind target regions with high affinity and specificity. It is preferred that the triplex forming molecules bind the target molecule with a k_d less than 10^{-6} , 10^{-8} , 10^{-10} , or 10^{-12} . Representative examples of how to make and use triplex forming molecules to bind a variety of different target molecules can be found in the following non-limiting list of U.S. Pat. Nos. 5,176,996, 5,645,985, 5,650,316, 5,683,874, 5,693,773, 5,834,185, 5,869,246, 5,874,566, and 5,962,426.

External guide sequences (EGSs) are molecules that bind a target nucleic acid molecule forming a complex, and this complex is recognized by RNase P, which cleaves the target molecule. EGSs can be designed to specifically target a RNA molecule of choice, RNase P aids in processing transfer RNA (tRNA) within a cell. Bacterial RNase P can be recruited to cleave virtually any RNA sequence by using an EGS that causes the target RNA:EGS complex to mimic the natural tRNA substrate. (WO 92/03566 by Yale, and Forster and Altman, *Science* 238:407-409 (1990)).

Similarly, eukaryotic EGS/RNase P-directed cleavage of RNA can be utilized to cleave desired targets within eukaryotic cells. (Yuan et al., *Proc. Natl. Acad. Sci. USA* 89:8006-8010 (1992); WO 93/22434 by Yale; WO 95/24489 by Yale; Yuan and Altman, *EMBO J* 14:159-168 (1995), and Carrara et al., *Proc. Natl. Acad. Sci. (USA)* 92:2627-2631 (1995)). Representative examples of how to make and use EGS molecules to facilitate cleavage of a variety of different target molecules be found in the following non-limiting list of U.S. Pat. Nos. 5,168,053, 5,624,824, 5,683,873, 5,728,521, 5,869,248, and 5,877,162.

4. Nucleic Acid Delivery

In the methods described above which include the administration and uptake of exogenous DNA into the cells of a subject (i.e., gene transduction or transfection), the disclosed nucleic acids can be in the form of naked DNA or RNA, or the nucleic acids can be in a vector for delivering the nucleic acids to the cells, whereby the antibody-encoding DNA fragment is under the transcriptional regulation of a promoter, as would be well understood by one of ordinary skill in the art. The vector can be a commercially available preparation, such as an adenovirus vector (Quantum Biotechnologies, Inc. (Laval, Quebec, Canada). Delivery of the nucleic acid or vector to cells can be via a variety of

mechanisms. As one example, delivery can be via a liposome, using commercially available liposome preparations such as LIPOFECTIN, LIPOFECTAMINE (GIBCO-BRL, Inc., Gaithersburg, Md.), SUPERFECT (Qiagen, Inc. Hilden, Germany) and TRANSFECTAM (Promega Biotech, Inc., Madison, Wis.), as well as other liposomes developed according to procedures standard in the art. In addition, the disclosed nucleic acid or vector can be delivered in vivo by electroporation, the technology for which is available from Genetronics, Inc. (San Diego, Calif.) as well as by means of a SONOPORATION machine (ImaRx Pharmaceutical Corp., Tucson, Ariz.).

As one example, vector delivery can be via a viral system, such as a retroviral vector system which can package a recombinant retroviral genome (see e.g., Pastan et al. *Proc. Natl. Acad. Sci. U.S.A.* 85:4486, 1988; Miller et al., *Mol. Cell. Biol.* 6:2895, 1986). The recombinant retrovirus can then be used to infect and thereby deliver to the infected cells nucleic acid encoding a broadly neutralizing antibody (or active fragment thereof). The exact method of introducing the altered nucleic acid into mammalian cells is, of course, not limited to the use of retroviral vectors. Other techniques are widely available for this procedure including the use of adenoviral vectors (Mitani et al., *Hum. Gene Ther.* 5:941-948, 1994), adeno-associated viral (AAV) vectors (Goodman et al., *Blood* 84:1492-1500, 1994), lentiviral vectors (Naidini et al., *Science* 272:263-267, 1996), pseudotyped retroviral vectors (Agrawal et al., *Exper. Hematol.* 24:738-747, 1996). Physical transduction techniques can also be used, such as liposome delivery and receptor-mediated and other endocytosis mechanisms (see, for example, Schwartzberger et al., *Blood* 87:472-478, 1996). This disclosed compositions and methods can be used in conjunction with any of these or other commonly used gene transfer methods.

As one example, if the antibody-encoding nucleic acid is delivered to the cells of a subject in an adenovirus vector, the dosage for administration of adenovirus to humans can range from about 10^7 to 10^9 plaque forming units (pfu) per injection but can be as high as 10^{12} pfu per injection (Crystal, *Hum. Gene Ther.* 8:985-1001, 1997; Alvarez and Curiel, *Hum. Gene Ther.* 8:597-613, 1997). A subject can receive a single injection, or, if additional injections are necessary, they can be repeated at six month intervals (or other appropriate time intervals, as determined by the skilled practitioner) for an indefinite period and/or until the efficacy of the treatment has been established.

Parenteral administration of the nucleic acid or vector, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. For additional discussion of suitable formulations and various routes of administration of therapeutic compounds, see, e.g., *Remington: The Science and Practice of Pharmacy* (19th ed.) ed. A. R. Gennaro, Mack Publishing Company, Easton, Pa. 1995.

5. Expression Systems

The nucleic acids that are delivered to cells typically contain expression controlling systems. For example, the inserted genes in viral and retroviral systems usually contain promoters, and/or enhancers to help control the expression of the desired gene product. A promoter is generally a sequence or sequences of DNA that function when in a relatively fixed location in regard to the transcription start

site. A promoter contains core elements required for basic interaction of RNA polymerase and transcription factors, and may contain upstream elements and response elements.

a) Viral Promoters and Enhancers

Preferred promoters controlling transcription from vectors in mammalian host cells may be obtained from various sources, for example, the genomes of viruses such as: polyoma, Simian Virus 40 (SV40), adenovirus, retroviruses, hepatitis-B virus and most preferably cytomegalovirus, or from heterologous mammalian promoters, e.g. beta actin promoter. The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment which also contains the SV40 viral origin of replication (Fiers et al., *Nature*, 273: 113 (1978)). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a HindIII E restriction fragment Greenway, P. J. et al., *Gene* 18: 355-360 (1982)). Of course, promoters from the host cell or related species also are useful herein.

Enhancer generally refers to a sequence of DNA that functions at no fixed distance from the transcription start site and can be either 5' (Laimins L. et al., *Proc. Natl. Acad. Sci.* 78: 993 (1981)) or 3' (Lusky, M. L., et al., *Mol. Cell Bio.* 3: 1108 (1983)) to the transcription unit. Furthermore, enhancers can be within an intron (Banerji, J. L. et al., *Cell* 33: 729 (1983)) as well as within the coding sequence itself (Osborne, T. F., et al., *Mol. Cell Bio.* 4: 1293 (1984)). They are usually between 10 and 300 bp in length, and they function in cis. Enhancers function to increase transcription from nearby promoters. Enhancers also often contain response elements that mediate the regulation of transcription. Promoters can also contain response elements that mediate the regulation of transcription. Enhancers often determine the regulation of expression of a gene. While many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin), typically one will use an enhancer from a eukaryotic cell virus for general expression. Preferred examples are the SV40 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

The promoter and/or enhancer may be specifically activated either by light or specific chemical events which trigger their function. Systems can be regulated by reagents such as tetracycline and dexamethasone. There are also ways to enhance viral vector gene expression by exposure to irradiation, such as gamma irradiation, or alkylating chemotherapy drugs.

In certain embodiments the promoter and/or enhancer region can act as a constitutive promoter and/or enhancer to maximize expression of the region of the transcription unit to be transcribed. In certain constructs the promoter and/or enhancer region be active in all eukaryotic cell types, even if it is only expressed in a particular type of cell at a particular time. A preferred promoter of this type is the CMV promoter (650 bases). Other preferred promoters are SV40 promoters, cytomegalovirus (full length promoter), and retroviral vector LTR.

It has been shown that all specific regulatory elements can be cloned and used to construct expression vectors that are selectively expressed in specific cell types such as melanoma cells. The glial fibrillary acetic protein (GFAP) promoter has been used to selectively express genes in cells of glial origin.

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human or nucleated cells) may also contain sequences necessary for the termination of

transcription which may affect mRNA expression. These regions are transcribed as polyadenylated segments in the untranslated portion of the mRNA encoding tissue factor protein. The 3' untranslated regions also include transcription termination sites. It is preferred that the transcription unit also contains a polyadenylation region. One benefit of this region is that it increases the likelihood that the transcribed unit will be processed and transported like mRNA. The identification and use of polyadenylation signals in expression constructs is well established. It is preferred that homologous polyadenylation signals be used in the transgene constructs. In certain transcription units, the polyadenylation region is derived from the SV40 early polyadenylation signal and consists of about 400 bases. It is also preferred that the transcribed units contain other standard sequences alone or in combination with the above sequences improve expression from, or stability of, the construct.

b) Markers

The viral vectors can include nucleic acid sequence encoding a marker product. This marker product is used to determine if the gene has been delivered to the cell and once delivered is being expressed. Preferred marker genes are the *E. Coli* lacZ gene, which encodes β -galactosidase, and green fluorescent protein.

In some embodiments the marker may be a selectable marker. Examples of suitable selectable markers for mammalian cells are dihydrofolate reductase (DHFR), thymidine kinase, neomycin, neomycin analog G418, hygromycin, and puromycin. When such selectable markers are successfully transferred into a mammalian host cell, the transformed mammalian host cell can survive if placed under selective pressure. There are two widely used distinct categories of selective regimes. The first category is based on a cell's metabolism and the use of a mutant cell line which lacks the ability to grow independent of a supplemented media. Two examples are: CHO DHFR-cells and mouse LTK-cells. These cells lack the ability to grow without the addition of such nutrients as thymidine or hypoxanthine. Because these cells lack certain genes necessary for a complete nucleotide synthesis pathway, they cannot survive unless the missing nucleotides are provided in a supplemented media. An alternative to supplementing the media is to introduce an intact DHFR or TK gene into cells lacking the respective genes, thus altering their growth requirements. Individual cells which were not transformed with the DHFR or TK gene will not be capable of survival in non-supplemented media.

The second category is dominant selection which refers to a selection scheme used in any cell type and does not require the use of a mutant cell line. These schemes typically use a drug to arrest growth of a host cell. Those cells which have a novel gene would express a protein conveying drug resistance and would survive the selection. Examples of such dominant selection use the drugs neomycin, (Southern P. and Berg, P., *J. Molec. Appl. Genet.* 1: 327 (1982)), mycophenolic acid, (Mulligan, R. C. and Berg, P. *Science* 209: 1422 (1980)) or hygromycin, (Sugden, B. et al., *Mol. Cell. Biol.* 5: 410-413 (1985)). The three examples employ bacterial genes under eukaryotic control to convey resistance to the appropriate drug G418 or neomycin (geneticin), xgpt (mycophenolic acid) or hygromycin, respectively. Others include the neomycin analog G418 and puramycin.

6. Peptides

a) Protein Variants

As discussed herein there are numerous variants of the isolated D4 desaturase and D5 elongase proteins that are known and herein contemplated. In addition, to the known

functional the isolated D4 desaturase and D5 elongase strain variants there are derivatives of the isolated D4 desaturase and D5 elongase proteins which also function in the disclosed methods and compositions. Protein variants and derivatives are well understood to those of skill in the art and in can involve amino acid sequence modifications. For example, amino acid sequence modifications typically fall into one or more of three classes: substitutional, insertional or deletional variants. Insertions include amino and/or carboxyl terminal fusions as well as intrasequence insertions of single or multiple amino acid residues. Insertions ordinarily will be smaller insertions than those of amino or carboxyl terminal fusions, for example, on the order of one to four residues. Immunogenic fusion protein derivatives, such as those described in the examples, are made by fusing a polypeptide sufficiently large to confer immunogenicity to the target sequence by cross-linking in vitro or by recombinant cell culture transformed with DNA encoding the fusion. Deletions are characterized by the removal of one or more amino acid residues from the protein sequence. Typically, no more than about from 2 to 6 residues are deleted at any one site within the protein molecule. These variants ordinarily are prepared by site specific mutagenesis of nucleotides in the DNA encoding the protein, thereby producing DNA encoding the variant, and thereafter expressing the DNA in recombinant cell culture. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence are well known, for example M13 primer mutagenesis and PCR mutagenesis. Amino acid substitutions are typically of single residues, but can occur at a number of different locations at once; insertions usually will be on the order of about from 1 to 10 amino acid residues; and deletions will range about from 1 to 30 residues. Deletions or insertions preferably are made in adjacent pairs, i.e. a deletion of 2 residues or insertion of 2 residues. Substitutions, deletions, insertions or any combination thereof may be combined to arrive at a final construct. The mutations must not place the sequence out of reading frame and preferably will not create complementary regions that could produce secondary mRNA structure. Substitutional variants are those in which at least one residue has been removed and a different residue inserted in its place. Such substitutions generally are made in accordance with the following Tables 2 and 3 and are referred to as conservative substitutions.

Substantial changes in function or immunological identity are made by selecting substitutions that are less conservative than those in Table 3, i.e., selecting residues that differ more significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site or (c) the bulk of the side chain. The substitutions which in general are expected to produce the greatest changes in the protein properties will be those in which (a) a hydrophilic residue, e.g. seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g. leucyl, isoleucyl, phenylalanyl, valyl or amyl; (b) a cysteine or proline is substituted for (or by) an other residue; (c) a residue having an electropositive side chain, e.g., lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g., glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g., phenylalanine, is substituted for (or by) one not having a side chain, e.g., glycine, in this case, (e) by increasing the number of sites for sulfation and/or glycosylation.

TABLE 2

Amino Acid Abbreviations		
Amino Acid	Abbreviations	
alanine	Ala	A
arginine	Arg	R
asparagine	Asn	N
aspartic acid	Asp	D
cysteine	Cys	C
glutamic acid	Glu	E
glutamine	Gln	K
glycine	Gly	G
histidine	His	H
isoleucine	Ile	I
leucine	Leu	L
lysine	Lys	K
phenylalanine	Phe	F
proline	Pro	P
serine	Ser	S
threonine	Thr	T
tyrosine	Tyr	Y
tryptophan	Trp	W
valine	Val	V
methionine	Met	M

For example, the replacement of one amino acid residue with another that is biologically and/or chemically similar is known to those skilled in the art as a conservative substitution. For example, a conservative substitution would be replacing one hydrophobic residue for another, or one polar residue for another. The substitutions include combinations such as, for example, Gly, Ala; Val, Ile, Leu; Asp, Glu; Asn, Gln; Ser, Thr; Lys, Arg; and Phe, Tyr. Such conservatively substituted variations of each explicitly disclosed sequence are included within the mosaic polypeptides provided herein.

Substitutional or deletional mutagenesis can be employed to insert sites for N-glycosylation (Asn-X-Thr/Ser) or O-glycosylation (Ser or Thr). Deletions of cysteine or other labile residues also may be desirable. Deletions or substitutions of potential proteolysis sites, e.g. Arg, is accomplished for example by deleting one of the basic residues or substituting one by glutaminyl or histidyl residues.

TABLE 3

Amino Acid Substitutions	
Original Residue	Exemplary Conservative Substitutions, others are known in the art.
	Ala; ser
	Arg; lys, gln
	Asn; gln; his
	Asp; glu
	Cys; ser
	Gln; asn, lys
	Glu; asp
	Gly; pro
	His; asn; gln
	Ile; leu; val
	Leu; ile; val
	Lys; arg; gln;
	Met; Leu; ile
	Phe; met; leu; tyr
	Ser; thr
	Thr; ser
	Trp; tyr
	Tyr; trp; phe
	Val; ile; leu

Certain post-translational derivatizations are the result of the action of recombinant host cells on the expressed polypeptide. Glutaminyl and asparaginyl residues are frequently

post-translationally deamidated to the corresponding glutamyl and aspartyl residues. Alternatively, these residues are deamidated under mildly acidic conditions. Other post-translational modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the o-amino groups of lysine, arginine, and histidine side chains (T. E. Creighton, *Proteins: Structure and Molecular Properties*, W. H. Freeman & Co., San Francisco pp 79-86 [1983]), acetylation of the N-terminal amine and, in some instances, amidation of the C-terminal carboxyl.

It is understood that one way to define the variants and derivatives of the disclosed proteins herein is through defining the variants and derivatives in terms of homology/identity to specific known sequences. For example, SEQ ID NO: 15 sets forth a particular sequence of a D5 elongase and SEQ ID NO:26 sets forth a particular sequence of a D4 desaturase protein. Specifically disclosed are variants of these and other proteins herein disclosed which have at least, 70% or 75% or 80% or 85% or 90% or 95% homology to the stated sequence. Those of skill in the art readily understand how to determine the homology of two proteins. For example, the homology can be calculated after aligning the two sequences so that the homology is at its highest level. Specifically disclosed are sequences having greater than 96% identity and 89% identity to SEQ ID NOs: 15 and 26.

Another way of calculating homology can be performed by published algorithms. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci. U.S.A.* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by inspection.

The same types of homology can be obtained for nucleic acids by for example the algorithms disclosed in Zuker, *M. Science* 244:48-52, 1989, Jaeger et al. *Proc. Natl. Acad. Sci. USA* 86:7706-7710, 1989, Jaeger et al. *Methods Enzymol.* 183:281-306, 1989 which are herein incorporated by reference for at least material related to nucleic acid alignment.

It is understood that the description of conservative mutations and homology can be combined together in any combination, such as embodiments that have at least 70% homology to a particular sequence wherein the variants are conservative mutations.

As this specification discusses various proteins and protein sequences it is understood that the nucleic acids that can encode those protein sequences are also disclosed. This would include all degenerate sequences related to a specific protein sequence, i.e. all nucleic acids having a sequence that encodes one particular protein sequence as well as all nucleic acids, including degenerate nucleic acids, encoding the disclosed variants and derivatives of the protein sequences. Thus, while each particular nucleic acid sequence may not be written out herein, it is understood that each and every sequence is in fact disclosed and described herein through the disclosed protein sequence. For example, one of the many nucleic acid sequences that can encode the protein sequence set forth in SEQ ID NO:15 is set forth in SEQ ID NO:14. It is also understood that while no amino acid sequence indicates what particular DNA sequence encodes that protein within an organism, where particular variants of a disclosed protein are disclosed herein, the

known nucleic acid sequence that encodes that protein in the particular strain from which that protein arises is also known and herein disclosed and described.

It is understood that there are numerous amino acid and peptide analogs which can be incorporated into the disclosed compositions. For example, there are numerous D amino acids or amino acids which have a different functional substituent then the amino acids shown in Table 2 and Table 3. The opposite stereo isomers of naturally occurring peptides are disclosed, as well as the stereo isomers of peptide analogs. These amino acids can readily be incorporated into polypeptide chains by charging tRNA molecules with the amino acid of choice and engineering genetic constructs that utilize, for example, amber codons, to insert the analog amino acid into a peptide chain in a site specific way (Thorson et al., *Methods in Molec. Biol.* 77:43-73 (1991), Zoller, *Current Opinion in Biotechnology*, 3:348-354 (1992); Ibba, *Biotechnology & Genetic Engineering Reviews* 13:197-216 (1995), Cahill et al, *TIBS*, 14(10):400-403 (1989); Benner, *TIB Tech*, 12:158-163 (1994); Ibba and Hennecke, *Bio/technology*, 12:678-682 (1994) all of which are herein incorporated by reference at least for material related to amino acid analogs).

Molecules can be produced that resemble peptides, but which are not connected via a natural peptide linkage. For example, linkages for amino acids or amino acid analogs can include CH_2NH —, $\text{—CH}_2\text{S}$ —, $\text{—CH}_2\text{—CH}_2\text{—}$, —CH=CH— (cis and trans), $\text{—COOH}_2\text{—}$, $\text{—CH(OH)CH}_2\text{—}$, and $\text{—CHH}_2\text{SO—}$ (These and others can be found in Spatola, F. in *Chemistry and Biochemistry of Amino Acids, Peptides, and Proteins*, B. Weinstein, eds., Marcel Dekker, New York, p. 267 (1983); Spatola, A. F., *Vega Data* (March 1983), Vol. 1, Issue 3, *Peptide Backbone Modifications* (general review); Morley, *Trends Pharm. Sci* (1980) pp. 463-468; Hudson, D. et al., *Int Pept Prot Res* 14:177-185 (1979) ($\text{—CH}_2\text{NH—}$, $\text{CH}_2\text{CH}_2\text{—}$); Spatola et al. *Life Sci* 38:1243-1249 (1986) ($\text{—CHH}_2\text{—S}$); Hann J. *Chem. Soc Perkin Trans. I* 307-314 (1982) (—CH—CH— , cis and trans); Almquist et al. *J. Med. Chem.* 23:1392-1398 (1980) ($\text{—COCH}_2\text{—}$); Jennings-White et al. *Tetrahedron Lett* 23:2533 (1982) ($\text{—COCH}_2\text{—}$); Szelke et al, *European Appln, EP 45665 CA* (1982): 97:39405 (1982) ($\text{—CH(OH)CH}_2\text{—}$); Holladay et al. *Tetrahedron. Lett* 24:4401-4404 (1983) ($\text{—C(OH)CH}_2\text{—}$); and Hruby *Life Sci* 31:189-199 (1982) ($\text{—CH}_2\text{—S—}$); each of which is incorporated herein by reference. A particularly preferred non-peptide linkage is $\text{—CH}_2\text{NH—}$. It is understood that peptide analogs can have more than one atom between the bond atoms, such as b-alanine, g-aminobutyric acid, and the like.

Amino acid analogs and analogs and peptide analogs often have enhanced or desirable properties, such as, more economical production, greater chemical stability, enhanced pharmacological properties (half-life, absorption, potency, efficacy, etc.), altered specificity (e.g., a broad-spectrum of biological activities), reduced antigenicity, and others.

D-amino acids can be used to generate more stable peptides, because D amino acids are not recognized by peptidases and such. Systematic substitution of one or more amino acids of a consensus sequence with a D-amino acid of the same type (e.g. D-lysine in place of L-lysine) can be used to generate more stable peptides, Cysteine residues can be used to cyclize or attach two or more peptides together. This can be beneficial to constrain peptides into particular conformations. (Rizo and Gierasch *Ann. Rev. Biochem.* 61:387 (1992), incorporated herein by reference).

7. Antibodies

(1) Antibodies Generally

The term "antibodies" is used herein in a broad sense and includes both polyclonal and monoclonal antibodies. In addition to intact immunoglobulin molecules, also included in the term "antibodies" are fragments or polymers of those immunoglobulin molecules, and human or humanized versions of immunoglobulin molecules or fragments thereof, as long as they are chosen for their ability to interact with the isolated D4 desaturases and D5 elongases such that they can be identified, bound, purified, or have altered activity. Antibodies that bind the disclosed regions of the D4 desaturases and D5 elongases are also disclosed. The antibodies can be tested for their desired activity using the in vitro assays described herein, or by analogous methods, after which their in vivo therapeutic and/or prophylactic activities are tested according to known clinical testing methods.

The term "monoclonal antibody" as used herein refers to an antibody obtained from a substantially homogeneous population of antibodies, the individual antibodies within the population are identical except for possible naturally occurring mutations that may be present in a small subset of the antibody molecules. The monoclonal antibodies herein specifically include "chimeric" antibodies in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, as long as they exhibit the desired antagonistic activity (See, U.S. Pat. No. 4,816,567 and Morrison et al., *Proc. Natl. Acad. Sci. USA*, 81:6851-6855 (1984)).

The disclosed monoclonal antibodies can be made using any procedure which produces mono clonal antibodies. For example, disclosed monoclonal antibodies can be prepared using hybridoma methods, such as those described by Kohler and Milstein, *Nature*, 256:495 (1975). In a hybridoma method, a mouse or other appropriate host animal is typically immunized with an immunizing agent to elicit lymphocytes that produce or are capable of producing antibodies that will specifically bind to the immunizing agent. Alternatively, the lymphocytes may be immunized in vitro, e.g., using, the HIV Env-CD4-co-receptor complexes described herein.

The monoclonal antibodies may also be made by recombinant DNA methods, such as those described in U.S. Pat. No. 4,816,567 (Cabilly et al.). DNA encoding the disclosed monoclonal antibodies can be readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of murine antibodies). Libraries of antibodies or active antibody fragments can also be generated and screened using phage display techniques, e.g., as described in U.S. Pat. No. 5,804,440 to Burton et al. and U.S. Pat. No. 6,096,411 to Barbas et al.

In vitro methods are also suitable for preparing monovalent antibodies. Digestion of antibodies to produce fragments thereof, particularly, Fab fragments, can be accomplished using routine techniques known in the art. For instance, digestion can be performed using papain. Examples of papain digestion are described in WO 94/29348 published Dec. 22, 1994 and U.S. Pat. No. 4,342,566. Papain digestion of antibodies typically produces two identical antigen binding fragments, called Fab fragments, each with

a single antigen binding site, and a residual Fe fragment. Pepsin treatment yields a fragment that has two antigen combining sites and is still capable of cross-linking antigen.

The fragments, whether attached to other sequences or not, can also include insertions, deletions, substitutions, or other selected modifications of particular regions or specific amino acids residues, provided the activity of the antibody or antibody fragment is not significantly altered or impaired compared to the non-modified antibody or antibody fragment. These modifications can provide for some additional property, such as to remove/add amino acids capable of disulfide bonding, to increase its bio-longevity, to alter its secretory characteristics, etc. In any case, the antibody or antibody fragment must possess a bioactive property, such as specific binding to its cognate antigen. Functional or active regions of the antibody or antibody fragment may be identified by mutagenesis of a specific region of the protein, followed by expression and testing of the expressed polypeptide. Such methods are readily apparent to a skilled practitioner in the art and can include site-specific mutagenesis of the nucleic acid encoding the antibody or antibody fragment. (Zoller, M. S. *Curr. Opin. Biotechnol.* 3:348-354, 1992).

As used herein, the term "antibody" or "antibodies" can also refer to a human antibody and/or a humanized antibody. Many non-human antibodies (e.g., those derived from mice, rats, or rabbits) are naturally antigenic in humans, and thus can give rise to undesirable immune responses when administered to humans. Therefore, the use of human or humanized antibodies in the methods serves to lessen the chance that an antibody administered to a human will evoke an undesirable immune response.

(2) Human Antibodies

The disclosed human antibodies can be prepared using any technique. Examples of techniques for human monoclonal antibody production include those described by Cole et al. (*Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, p. 77, 1985) and by Boerner et al. (*J. Immunol.*, 147(1):86-95, 1991). Human antibodies (and fragments thereof) can also be produced using Phage display libraries (Hoogenboom et al., *J. Mol. Biol.*, 227:381, 1991; Marks et al., *J. Mol. Biol.*, 222:581, 1991).

The disclosed human antibodies can also be obtained from transgenic animals. For example, transgenic, mutant mice that are capable of producing a full repertoire of human antibodies, in response to immunization, have been described (see, e.g., Jakobovits et al., *Proc. Natl. Acad. Sci. USA*, 90:2551-255 (1993); Jakobovits et al., *Nature*, 362:255-258 (1993); Bruggermann et al., *Year in Immunol.*, 7:33 (1993)). Specifically, the homozygous deletion of the antibody heavy chain joining region (J(H)) gene in these chimeric and germ-line mutant mice results in complete inhibition of endogenous antibody production, and the successful transfer of the human germ-line antibody gene array into such germ-line mutant mice results in the production of human antibodies upon antigen challenge. Antibodies having the desired activity are selected using Env-CD4-co-receptor complexes as described herein.

(3) Humanized Antibodies

Antibody humanization techniques generally involve the use of recombinant DNA technology to manipulate the DNA sequence encoding one or more polypeptide chains of an antibody molecule. Accordingly, a humanized form of a non-human antibody (or a fragment thereof) is a chimeric antibody or antibody chain (or a fragment thereof, such as an Fv, Fab, Fab', or other antigen-binding portion of an antibody) which contains a portion of an antigen binding site

from a non-human (donor) antibody integrated into the framework of a human (recipient) antibody.

To generate a humanized antibody, residues from one or more complementarity determining regions (CDRs) of a recipient (human) antibody molecule are replaced by residues from one or more CDRs of a donor (non-human) antibody molecule that is known to have desired antigen binding characteristics (e.g., a certain level of specificity and affinity for the target antigen). In some instances, Fv framework (FR) residues of the human antibody are replaced by corresponding non-human residues. Humanized antibodies may also contain residues which are found neither in the recipient antibody nor in the imported CDR or framework sequences. Generally, a humanized antibody has one or more amino acid residues introduced into it from a source which is non-human. In practice, humanized antibodies are typically human antibodies in which some CDR residues and possibly some FR residues are substituted by residues from analogous sites in rodent antibodies. Humanized antibodies generally contain at least a portion of an antibody constant region (Fc), typically that of a human antibody (Jones et al., *Nature*, 321:522-525 (1986), Reichmann et al., *Nature*, 332:323-327 (1988), and Presta, *Curr. Opin. Struct. Biol.*, 2:593-596 (1992)).

Methods for humanizing non-human antibodies are well known in the art. For example, humanized antibodies can be generated according to the methods of Winter and coworkers (Jones et al., *Nature*, 321:522-525 (1986), Reichmann et al., *Nature*, 332:323-327 (1988), Verhoeyen et al., *Science*, 239:1534-1536 (1988)), by substituting rodent CDRs or CDR sequences for the corresponding sequences of a human antibody. Methods that can be used to produce humanized antibodies are also described in U.S. Pat. No. 4,816,567 (Cabilly et al.), U.S. Pat. No. 5,565,332 (Hoogenboom et al.), U.S. Pat. No. 5,721,367 (Kay et al.), U.S. Pat. No. 5,837,243 (Deo et al.), U.S. Pat. No. 5,939,598 (Kucherlapati et al.), U.S. Pat. No. 6,130,364 (Jakobovits et al.), and U.S. Pat. No. 6,180,377 (Morgan et al.).

(4) Administration of Antibodies

Administration of the antibodies can be done as disclosed herein. Nucleic acid approaches for antibody delivery also exist. The antibodies and antibody fragments can also be administered to patients or subjects or cells as a nucleic acid preparation (e.g., DNA or RNA) that encodes the antibody or antibody fragment, such that the patient's or subject's own cells take up the nucleic acid and produce and secrete the encoded antibody or antibody fragment. The delivery of the nucleic acid can be by any means, as disclosed herein, for example.

8. Pharmaceutical Carriers/Delivery of Pharmaceutical Products

As described above, the compositions can also be administered in vivo in a pharmaceutically acceptable carrier. By "pharmaceutically acceptable" is meant a material that is not biologically or otherwise undesirable, i.e., the material may be administered to a subject, along with the nucleic acid or vector, without causing any undesirable biological effects or interacting in a deleterious manner with any of the other components of the pharmaceutical composition in which it is contained. The carrier would naturally be selected to minimize any degradation of the active ingredient and to minimize any adverse side effects in the subject, as would be well known to one of skill in the art.

The compositions may be administered orally, parenterally (e.g., intravenously), by intramuscular injection, by intraperitoneal injection, transdermally, extracorporeally, topically or the like, including topical intranasal adminis-

tration or administration by inhalant. As used herein, "topical intranasal administration" means delivery of the compositions into the nose and nasal passages through one or both of the nares and can comprise delivery by a spraying mechanism or droplet mechanism, or through aerosolization of the nucleic acid or vector. Administration of the compositions by inhalant can be through the nose or mouth via delivery by a spraying or droplet mechanism. Delivery can also be directly to any area of the respiratory system (e.g., lungs) via intubation. The exact amount of the compositions required will vary from subject to subject, depending on the species, age, weight and general condition of the subject, the severity of the allergic disorder being treated, the particular nucleic acid or vector used, its mode of administration and the like. Thus, it is not possible to specify an exact amount for every composition. However, an appropriate amount can be determined by one of ordinary skill in the art using only routine experimentation given the teachings herein.

Parenteral administration of the composition, if used, is generally characterized by injection. Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant dosage is maintained. See, e.g., U.S. Pat. No. 3,610,795, which is incorporated by reference herein.

The materials may be in solution, suspension (for example, incorporated into microparticles, liposomes, or cells). These may be targeted to a particular cell type via antibodies, receptors, or receptor ligands. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Senter, et al., *Bioconjugate Chem.*, 2:447-451, (1991); Bagshawe, K. D., *Br. J. Cancer*, 60:275-281, (1989); Bagshawe, et al., *Br. J. Cancer*, 58:700-703, (1988); Senter, et al., *Bioconjugate Chem.*, 4:3-9, (1993); Battelli, et al., *Cancer Immunol. Immunother.*, 35:421-425, (1992); Pietersz and McKenzie, *Immunolog. Reviews*, 129:57-80, (1992); and Roffler, et al., *Biochem. Pharmacol.*, 42:2062-2065, (1991)). Vehicles such as "stealth" and other antibody conjugated liposomes (including lipid mediated drug targeting to colonic carcinoma), receptor mediated targeting of DNA through cell specific ligands, lymphocyte directed tumor targeting, and highly specific therapeutic retroviral targeting of murine glioma cells in vivo. The following references are examples of the use of this technology to target specific proteins to tumor tissue (Hughes et al., *Cancer Research*, 49:6214-6220, (1989); and Litzinger and Huang, *Biochimica et Biophysica Acta*, 1104:179-187, (1992)). In general, receptors are involved in pathways of endocytosis, either constitutive or ligand induced. These receptors cluster in clathrin-coated pits, enter the cell via clathrin-coated vesicles, pass through an acidified endosome in which the receptors are sorted, and then either recycle to the cell surface, become stored intracellularly, or are degraded in lysosomes. The internalization pathways serve a variety of functions, such as nutrient uptake, removal of activated proteins, clearance of macromolecules, opportunistic entry of viruses and toxins, dissociation and degradation of ligand, and receptor-level regulation. Many receptors follow more than one intracellular pathway, depending on the cell type, receptor concentration, type of ligand, ligand valency, and ligand concentration. Molecular and cellular mechanisms of receptor-mediated endocytosis has been reviewed (Brown and Greene, *DNA and Cell Biology* 10:6, 399-409 (1991)).

a) Pharmaceutically Acceptable Carriers

The compositions, including antibodies, can be used therapeutically in combination with a pharmaceutically acceptable carrier.

Suitable carriers and their formulations are described in *Remington: The Science and Practice of Pharmacy* (19th ed.) ed. A. R. Gennaro, Mack Publishing Company, Easton, Pa. 1995. Typically, an appropriate amount of a pharmaceutically-acceptable salt is used in the formulation to render the formulation isotonic. Examples of the pharmaceutically-acceptable carrier include, but are not limited to, saline, Ringer's solution and dextrose solution. The pH of the solution is preferably from about 5 to about 8, and more preferably from about 7 to about 7.5. Further carriers include sustained release preparations such as semipermeable matrices of solid hydrophobic polymers containing the antibody, which matrices are in the form of shaped articles, e.g., films, liposomes or microparticles. It will be apparent to those persons skilled in the art that certain carriers may be more preferable depending upon, for instance, the route of administration and concentration of composition being administered.

Pharmaceutical carriers are known to those skilled in the art. These most typically would be standard carriers for administration of drugs to humans, including solutions such as sterile water, saline, and buffered solutions at physiological pH. The compositions can be administered intramuscularly or subcutaneously. Other compounds will be administered according to standard procedures used by those skilled in the art.

Pharmaceutical compositions may include carriers, thickeners, diluents, buffers, preservatives, surface active agents and the like in addition to the molecule of choice. Pharmaceutical compositions may also include one or more active ingredients such as antimicrobial agents, antiinflammatory agents, anesthetics, and the like.

The pharmaceutical composition may be administered in a number of ways depending on whether local or systemic treatment is desired, and on the area to be treated. Administration may be topically (including ophthalmically, vaginally, rectally, intranasally), orally, by inhalation, or parenterally, for example by intravenous drip, subcutaneous, intraperitoneal or intramuscular injection. The disclosed antibodies can be administered intravenously, intraperitoneally, intramuscularly, subcutaneously, intracavity, or transdermally.

Preparations for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's, or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers (such as those based on Ringer's dextrose), and the like. Preservatives and other additives may also be present such as, for example, antimicrobials, anti-oxidants, chelating agents, and inert gases and the like.

Formulations for topical administration may include ointments, lotions, creams, gels, drops, suppositories, sprays, liquids and powders. Conventional pharmaceutical carriers, aqueous, powder or oily bases, thickeners and the like may be necessary or desirable.

Compositions for oral administration include powders or granules, suspensions or solutions in water or non-aqueous media, capsules, sachets, or tablets. Thickeners, flavorings, diluents, emulsifiers, dispersing aids or binders may be desirable.

Some of the compositions may potentially be administered as a pharmaceutically acceptable acid- or base-addition salt, formed by reaction with inorganic acids such as hydrochloric acid, hydrobromic acid, perchloric acid, nitric acid, thiocyanic acid, sulfuric acid, and phosphoric acid, and organic acids such as formic acid, acetic acid, propionic acid, glycolic acid, lactic acid, pyruvic acid, oxalic acid, malonic acid, succinic acid, maleic acid, and fumaric acid, or by reaction with an inorganic base such as sodium hydroxide, ammonium hydroxide, potassium hydroxide, and organic bases such as mono-, di-, trialkyl and aryl amines and substituted ethanolamines.

b) Therapeutic Uses

Effective dosages and schedules for administering the compositions may be determined empirically, and making such determinations is within the skill in the art. The dosage ranges for the administration of the compositions are those large enough to produce the desired effect in which the symptoms disorder are effected. The dosage should not be so large as to cause adverse side effects, such as unwanted cross-reactions, anaphylactic reactions, and the like. Generally, the dosage will vary with the age, condition, sex and extent of the disease in the patient, route of administration, or whether other drugs are included in the regimen, and can be determined by one of skill in the art. The dosage can be adjusted by the individual physician in the event of any counterindications. Dosage can vary, and can be administered in one or more dose administrations daily, for one or several days. Guidance can be found in the literature for appropriate dosages for given classes of pharmaceutical products. For example, guidance in selecting appropriate doses for antibodies can be found in the literature on therapeutic uses of antibodies, e.g., *Handbook of Monoclonal Antibodies*, Ferrone et al., eds., Noyes Publications, Park Ridge, N.J., (1985) ch. 22 and pp. 303-357; Smith et al., *Antibodies in Human Diagnosis and Therapy*, Haber et al., eds., Raven Press, New York (1977) pp. 365-389. A typical daily dosage of the antibody used alone might range from about 1 µg/kg to up to 100 mg/kg of body weight or more per day, depending on the factors mentioned above.

Other molecules that interact with the isolated D4 desaturases and D5 elongases which do not have a specific pharmaceutical function but which may be used for tracking changes within cellular chromosomes or for the delivery of diagnostic tools for example can be delivered in ways similar to those described for the pharmaceutical products.

9. Chips and Micro Arrays

Disclosed are chips where at least one address is the sequences or part of the sequences set forth in any of the nucleic acid sequences disclosed herein. Also disclosed are chips where at least one address is the sequences or portion of sequences set forth in any of the peptide sequences disclosed herein.

Also disclosed are chips where at least one address is a variant of the sequences or part of the sequences set forth in any of the nucleic acid sequences disclosed herein. Also disclosed are chips where at least one address is a variant of the sequences or portion of sequences set forth in any of the peptide sequences disclosed herein.

10. Computer Readable Mediums

It is understood that the disclosed nucleic acids and proteins can be represented as a sequence consisting of the

nucleotides of amino acids. There are a variety of ways to display these sequences, for example the nucleotide guanosine can be represented by G or g. Likewise the amino acid valine can be represented by Val or V. Those of skill in the art understand how to display and express any nucleic acid or protein sequence in any of the variety of ways that exist, each of which is considered herein disclosed. Specifically contemplated herein is the display of these sequences on computer readable mediums, such as, commercially available floppy disks, tapes, chips, hard drives, compact disks, and video disks, or other computer readable mediums. Also disclosed are the binary code representations of the disclosed sequences. Those of skill in the art understand what computer readable mediums. Thus, computer readable mediums on which the nucleic acids or protein sequences are recorded, stored, or saved.

Disclosed are computer readable mediums comprising the sequences and information regarding the sequences set forth herein.

11. Compositions Identified by Screening with Disclosed Compositions Combinatorial Chemistry

a) Combinatorial Chemistry

The disclosed compositions can be used as targets for any combinatorial technique to identify molecules or macromolecular molecules that interact with the disclosed compositions in a desired way. The nucleic acids, peptides, and related molecules disclosed herein can be used as targets for the combinatorial approaches. Also disclosed are the compositions that are identified through combinatorial techniques or screening techniques in which the compositions disclosed in for example SEQ ID NOS:14, 15, 25, or 26 or portions thereof, are used as the target in a combinatorial or screening protocol.

It is understood that when using the disclosed compositions in combinatorial techniques or screening methods, molecules, such as macromolecular molecules, will be identified that have particular desired properties such as inhibition or stimulation or the target molecule's function. The molecules identified and isolated when using the disclosed compositions, such as, a fatty acid, are also disclosed. Thus, the products produced using the combinatorial or screening approaches that involve the disclosed compositions, such as, fatty acids, are also considered herein disclosed.

155. It is understood that the disclosed methods for identifying molecules that inhibit the interactions between, for example, the isolated D4 desaturases and the D5 don gases can be performed using high through put means. For example, putative inhibitors can be identified using Fluorescence Resonance Energy Transfer (FRET) to quickly identify interactions. The underlying theory of the techniques is that when two molecules are close in space, ie, interacting at a level beyond background, a signal is produced or a signal can be quenched. Then, a variety of experiments can be performed, including, for example, adding in a putative inhibitor. If the inhibitor competes with the interaction between the two signaling molecules, the signals will be removed from each other in space, and this will cause a decrease or an increase in the signal, depending on the type of signal used. This decrease or increasing signal can be correlated to the presence or absence of the putative inhibitor. Any signaling means can be used. For example, disclosed are methods of identifying an inhibitor of the interaction between any two of the disclosed molecules comprising, contacting a first molecule and a second molecule together in the presence of a putative inhibitor, wherein the first molecule or second molecule comprises a fluorescence donor, wherein the first or second molecule,

typically the molecule not comprising the donor, comprises a fluorescence acceptor; and measuring Fluorescence Resonance Energy Transfer (FRET), in the presence of the putative inhibitor and the in absence of the putative inhibitor, wherein a decrease in FRET in the presence of the putative inhibitor as compared to FRET measurement in its absence indicates the putative inhibitor inhibits binding between the two molecules. This type of method can be performed with a cell system as well.

Combinatorial chemistry includes but is not limited to all methods for isolating small molecules or macromolecules that are capable of binding either a small molecule or another macromolecule, typically in an iterative process. Proteins, oligonucleotides, and sugars are examples of macromolecules. For example, oligonucleotide molecules with a given function, catalytic or ligand-binding, can be isolated from a complex mixture of random oligonucleotides in what has been referred to as "in vitro genetics" (Szostak, *TIBS* 19:89, 1992). One synthesizes a large pool of molecules bearing random and defined sequences and subjects that complex mixture, for example, approximately 10^{15} individual sequences in 100 μ g of a 100 nucleotide RNA, to some selection and enrichment process. Through repeated cycles of affinity chromatography and PCR amplification of the molecules bound to the ligand on the column, Ellington and Szostak (1990) estimated that 1 in 10^{10} RNA molecules folded in such a way as to bind a small molecule dyes. DNA molecules with such ligand-binding behavior have been isolated as well (Ellington and Szostak, 1992; Bock et al, 1992). Techniques aimed at similar goals exist for small organic molecules, proteins, antibodies and other macromolecules known to those of skill in the art. Screening sets of molecules for a desired activity whether based on small organic libraries, oligonucleotides, or antibodies is broadly referred to as combinatorial chemistry. Combinatorial techniques are particularly suited for defining binding interactions between molecules and for isolating molecules that have a specific binding activity, often called aptamers when the macromolecules are nucleic acids.

There are a number of methods for isolating proteins which either have de novo activity or a modified activity. For example, phage display libraries have been used to isolate numerous peptides that interact with a specific target. (See for example, U.S. Pat. Nos. 6,031,071; 5,824,520; 5,596,079; and 5,565,332 which are herein incorporated by reference at least for their material related to phage display and methods relate to combinatorial chemistry)

A preferred method for isolating proteins that have a given function is described by Roberts and Szostak (Roberts R. W. and Szostak J. W, Proc. Natl. Acad. Sci. USA, 94(23)12997-302 (1997). This combinatorial chemistry method couples the functional power of proteins and the genetic power of nucleic acids. An RNA molecule is generated in which a puromycin molecule is covalently attached to the 3'-end of the RNA molecule. An in vitro translation of this modified RNA molecule causes the correct protein, encoded by the RNA to be translated. In addition, because of the attachment of the puromycin, a peptidyl acceptor which cannot be extended, the growing peptide chain is attached to the puromycin which is attached to the RNA. Thus, the protein molecule is attached to the genetic material that encodes it. Normal in vitro selection procedures can now be done to isolate functional peptides. Once the selection procedure for peptide function is complete traditional nucleic acid manipulation procedures are performed to amplify the nucleic acid that codes for the selected functional peptides. After amplification of the genetic material,

new RNA is transcribed with puromycin at the 3'-end, new peptide is translated and another functional round of selection is performed. Thus, protein selection can be performed in an iterative manner just like nucleic acid selection techniques. The peptide which is translated is controlled by the sequence of the RNA attached to the puromycin. This sequence can be anything from a random sequence engineered for optimum translation (i.e. no stop codons etc.) or it can be a degenerate sequence of a known RNA molecule to look for improved or altered function of a known peptide. The conditions for nucleic acid amplification and in vitro translation are well known to those of ordinary skill in the art and are preferably performed as in Roberts and Szostak (Roberts R. W. and Szostak J. W. *Proc. Natl. Acad. Sci. USA*, 94(23)12997-302 (1997)).

Another preferred method for combinatorial methods designed to isolate peptides is described in Cohen et al (Cohen B. A., et al., *Proc. Natl. Acad. Sci. USA* 95(24): 14272-7 (1998)). This method utilizes and modifies two-hybrid technology. Yeast two-hybrid systems are useful for the detection and analysis of protein:protein interactions. The two-hybrid system, initially described in the yeast *Saccharomyces cerevisiae*, is a powerful molecular genetic technique for identifying new regulatory molecules, specific to the protein of interest (Fields and Song, *Nature* 340:245-6 (1989)). Cohen et al., modified this technology so that novel interactions between synthetic or engineered peptide sequences could be identified which bind a molecule of choice. The benefit of this type of technology is that the selection is done in an intracellular environment. The method utilizes a library of peptide molecules that attached to an acidic activation domain. A peptide of choice, for example a portion of the isolated D4 desaturases or D5 elongases is attached to a DNA binding domain of a transcriptional activation protein, such as Gal 4. By performing the Two-hybrid technique on this type of system, molecules that bind the portion of the isolated D4 desaturases or D5 elongases can be identified.

Using methodology well known to those of skill in the art, in combination with various combinatorial libraries, one can isolate and characterize those small molecules or macromolecules, which bind to or interact with the desired target. The relative binding affinity of these compounds can be compared and optimum compounds identified using competitive binding studies, which are well known to those of skill in the art.

Techniques for making combinatorial libraries and screening combinatorial libraries to isolate molecules which bind a desired target are well known to those of skill in the art. Representative techniques and methods can be found in but are not limited to U.S. Pat. Nos. 5,084,824, 5,288,514, 5,449,754, 5,506,337, 5,539,083, 5,545,568, 5,556,762, 5,565,324, 5,565,332, 5,573,905, 5,618,825, 5,619,680, 5,627,210, 5,646,285, 5,663,046, 5,670,326, 5,677,195, 5,683,899, 5,688,696, 5,688,997, 5,698,685, 5,712,146, 5,721,099, 5,723,598, 5,741,713, 5,792,431, 5,807,683, 5,807,754, 5,821,130, 5,831,014, 5,834,195, 5,834,318, 5,834,588, 5,840,500, 5,847,150, 5,856,107, 5,856,496, 5,859,190, 5,864,010, 5,874,443, 5,877,214, 5,880,972, 5,886,126, 5,886,127, 5,891,737, 5,916,899, 5,919,955, 5,925,527, 5,939,268, 5,942,387, 5,945,070, 5,948,696, 5,958,702, 5,958,792, 5,962,337, 5,965,719, 5,972,719, 5,976,894, 5,980,704, 5,985,356, 5,999,086, 6,001,579, 6,004,617, 6,008,321, 6,017,768, 6,025,371, 6,030,917, 6,040,193, 6,045,671, 6,045,755, 6,060,596, and 6,061,636.

Combinatorial libraries can be made from a wide array of molecules using a number of different synthetic techniques.

For example, libraries containing fused 2,4-pyrimidinediones (U.S. Pat. No. 6,025,371) dihydrobenzopyrans (U.S. Pat. Nos. 6,017,768 and 5,821,130), amide alcohols (U.S. Pat. No. 5,976,894), hydroxy-amino acid amides (U.S. Pat. No. 5,972,719) carbohydrates (U.S. Pat. No. 5,965,719), 1,4-benzodiazepin-2,5-diones (U.S. Pat. No. 5,962,337), cyclics (U.S. Pat. No. 5,958,792), biaryl amino acid amides (U.S. Pat. No. 5,948,696), thiophenes (U.S. Pat. No. 5,942,387), tricyclic Tetrahydroquinolines (U.S. Pat. No. 5,925,527), benzofurans (U.S. Pat. No. 5,919,955), isoquinolines (U.S. Pat. No. 5,916,899), hydantoin and thiohydantoin (U.S. Pat. No. 5,859,190), indoles (U.S. Pat. No. 5,856,496), imidazol-pyrido-indole and imidazol-pyrido-benzothio-phenes (U.S. Pat. No. 5,856,107) substituted 2-methylene-2,3-dihydrothiazoles (U.S. Pat. No. 5,847,150), quinolines (U.S. Pat. No. 5,840,500), PNA (U.S. Pat. No. 5,831,014), containing tags (U.S. Pat. No. 5,721,099), polyketides (U.S. Pat. No. 5,712,146), morpholino-subunits (U.S. Pat. Nos. 5,698,685 and 5,506,337), sulfamides (U.S. Pat. No. 5,618,825), and benzodiazepines (U.S. Pat. No. 5,288,514).

As used herein combinatorial methods and libraries included traditional screening methods and libraries as well as methods and libraries used in iterative processes.

b) Computer Assisted Drug Design

The disclosed compositions can be used as targets for any molecular modeling technique to identify either the structure of the disclosed compositions or to identify potential or actual molecules, such as small molecules, which interact in a desired way with the disclosed compositions. The nucleic acids, peptides, and related molecules disclosed herein can be used as targets in any molecular modeling program or approach.

It is understood that when using the disclosed compositions in modeling techniques, molecules, such as macromolecular molecules, will be identified that have particular desired properties such as inhibition or stimulation or the target molecule's function. The molecules identified and isolated when using the disclosed compositions, such as, the isolated D4 desaturases or D5 elongases, are also disclosed. Thus, the products produced using the molecular modeling approaches that involve the disclosed compositions, such as, the isolated D4 desaturases or D5 elongases, are also considered herein disclosed.

Thus, one way to isolate molecules that bind a molecule of choice is through rational design. This is achieved through structural information and computer modeling. Computer modeling technology allows visualization of the three-dimensional atomic structure of a selected molecule and the rational design of new compounds that will interact with the molecule. The three-dimensional construct typically depends on data from x-ray crystallographic analyses or NMR imaging of the selected molecule. The molecular dynamics require force field data. The computer graphics systems enable prediction of how a new compound will link to the target molecule and allow experimental manipulation of the structures of the compound and target molecule to perfect binding specificity. Prediction of what the molecule-compound interaction will be when small changes are made in one or both requires molecular mechanics software and computationally intensive computers, usually coupled with user-friendly, menu-driven interfaces between the molecular design program and the user.

Examples of molecular modeling systems are the CHARMM and QUANTA programs. Polygen Corporation, Waltham, Mass. CHARMM performs the energy minimization and molecular dynamics functions. QUANTA performs the construction, graphic modeling and analysis of molecu-

lar structure. QUANTA allows interactive construction, modification, visualization, and analysis of the behavior of molecules with each other.

A number of articles review computer modeling of drugs interactive with specific proteins, such as Rotivinen, et al., 1988 *Acta Pharmaceutica Fennica* 97, 159-166; Ripka, *New Scientist* 54-57 (Jun. 16, 1988); McKinaly and Rossmann, 1989 *Annu. Rev. Pharmacol. Toxicol.* 29, 111-122; Perry and Davies, QSAR: Quantitative Structure-Activity Relationships in Drug Design pp. 189-193 (Alan R. Liss, Inc. 1989); Lewis and Dean, 1989 *Proc. R. Soc. Lond.* 236, 125-140 and 141-162; and, with respect to a model enzyme for nucleic acid components, Askew, et al., 1989 *J. Am. Chem. Soc.* 111, 1082-1090. Other computer programs that screen and graphically depict chemicals are available from companies such as BioDesign, Inc., Pasadena, Calif., Allelix, Inc, Mississauga, Ontario, Canada, and Hypercube, Inc., Cambridge, Ontario. Although these are primarily designed for application to drugs specific to particular proteins, they can be adapted to design of molecules specifically interacting with specific regions of DNA or RNA, once that region is identified.

Although described above with reference to design and generation of compounds which could alter binding, one could also screen libraries of known compounds, including natural products or synthetic chemicals, and biologically active materials, including proteins, for compounds which alter substrate binding or enzymatic activity.

12. Kits

Disclosed herein are kits that are drawn to reagents that can be used in practicing the methods disclosed herein. The kits can include any reagent or combination of reagent discussed herein or that would be understood to be required or beneficial in the practice of the disclosed methods. For example, the kits could include primers to perform the amplification reactions discussed in certain embodiments of the methods, as well as the buffers and enzymes required to use the primers as intended.

13. Compositions with Similar Functions

It is understood that the compositions disclosed herein have certain functions, such as enzymatic functions disclosed. Disclosed herein are certain structural requirements for performing the disclosed functions, and it is understood that there are a variety of structures which can perform the same function which are related to the disclosed structures, and that these structures will ultimately achieve the same result, for example stimulation or inhibition of the enzymatic function.

C. Methods of Making the Compositions

The compositions disclosed herein and the compositions necessary to perform the disclosed methods can be made using any method known to those of skill in the art for that particular reagent or compound unless otherwise specifically noted.

1. Nucleic Acid Synthesis

For example, the nucleic acids, such as, the oligonucleotides to be used as primers can be made using standard chemical synthesis methods or can be produced using enzymatic methods or any other known method. Such methods can range from standard enzymatic digestion followed by nucleotide fragment isolation (see for example, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Edition (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989) Chapters 5, 6) to purely synthetic methods, for example, by the cyanoethyl phosphoramidite method using a Milligen or Beckman System I Plus DNA synthesizer (for example, Model 8700 automated synthesizer of Milligen-

Biosearch, Burlington, Mass. or ABI Model 380B). Synthetic methods useful for making oligonucleotides are also described by Ikuta et al., *Ann. Rev. Biochem.* 53:323-356 (1984), (phosphotriester and phosphite-triester methods), and Narang et al., *Methods Enzymol.*, 65:610-620 (1980), (phosphotriester method). Protein nucleic acid molecules can be made using known methods such as those described by Nielsen et al., *Bioconjug. Chem.* 5:3-7 (1994).

2. Peptide Synthesis

One method of producing the disclosed proteins, such as SEQ ID NO:23, is to link two or more peptides or polypeptides together by protein chemistry techniques. For example, peptides or polypeptides can be chemically synthesized using currently available laboratory equipment using either Fmoc (9-fluorenylmethyloxycarbonyl) or Boc (tert-butyloxycarbonyl) chemistry (Applied Biosystems, Inc., Foster City, Calif.). One skilled in the art can readily appreciate that a peptide or polypeptide corresponding to the disclosed proteins, for example, can be synthesized by standard chemical reactions. For example, a peptide or polypeptide can be synthesized and not cleaved from its synthesis resin whereas the other fragment of a peptide or protein can be synthesized and subsequently cleaved from the resin, thereby exposing a terminal group which is functionally blocked on the other fragment. By peptide condensation reactions, these two fragments can be covalently joined via a peptide bond at their carboxyl and amino termini, respectively, to form an antibody, or fragment thereof. (Grant G A (1992) *Synthetic Peptides: A User Guide*. W.H. Freeman and Co., N.Y. (1992); Bodansky M and Trost B., Ed. (1993) *Principles of Peptide Synthesis*. Springer-Verlag Inc., NY (which is herein incorporated by reference at least for material related to peptide synthesis). Alternatively, the peptide or polypeptide is independently synthesized in vivo as described herein. Once isolated, these independent peptides or polypeptides may be linked to form a peptide or fragment thereof via similar peptide condensation reactions.

For example, enzymatic ligation of cloned or synthetic peptide segments allow relatively short peptide fragments to be joined to produce larger peptide fragments, polypeptides or whole protein domains (Abrahmsen L et al., *Biochemistry*, 30:4151 (1991)). Alternatively, native chemical ligation of synthetic peptides can be utilized to synthetically construct large peptides or polypeptides from shorter peptide fragments. This method consists of a two step chemical reaction (Dawson et al. *Synthesis of Proteins by Native Chemical Ligation*. *Science*, 266:776-779 (1994)). The first step is the chemoselective reaction of an unprotected synthetic peptide-thioester with another unprotected peptide segment containing an amino-terminal Cys residue to give a thioester-linked intermediate as the initial covalent product. Without a change in the reaction conditions, this intermediate undergoes spontaneous, rapid intramolecular reaction to form a native peptide bond at the ligation site (Baggiolini M et al. (1992) *FEBS Lett.* 307:97-101; Clark-Lewis I et al., *J. Biol. Chem.* 269:16075 (1994); Clark-Lewis et al., *Biochemistry*, 30:3128 (1991); Rajarathnam K et al., *Biochemistry* 33:6623-30 (1994)).

Alternatively, unprotected peptide segments are chemically linked where the bond formed between the peptide segments as a result of the chemical ligation is an unnatural (non-peptide) bond (Schnolzer, M et al. *Science*, 256:221 (1992)). This technique has been used to synthesize analogs of protein domains as well as large amounts of relatively pure proteins with full biological activity (de Lisle Milton R C et al., *Techniques in Protein Chemistry IV*. Academic Press, New York, pp. 257-267 (1992)).

3. Process Claims for Making the Compositions

Disclosed are processes for making the compositions as well as making the intermediates leading to the compositions. For example, disclosed are nucleic acids in SEQ ID NOs:14 and 25. There are a variety of methods that can be used for making these compositions, such as synthetic chemical methods and standard molecular biology methods. It is understood that the methods of making these and the other disclosed compositions are specifically disclosed.

Disclosed are nucleic acid molecules produced by the process comprising linking in an operative way a nucleic acid comprising the sequence set forth in SEQ ID NO:14 and 25, for example, and a sequence controlling the expression of the nucleic acid.

Also disclosed are nucleic acid molecules produced by the process comprising linking in an operative way a nucleic acid molecule comprising a sequence having 80% identity to a sequence set forth in SEQ ID NO:14 and 25, for example, and a sequence controlling the expression of the nucleic acid.

Disclosed are nucleic acid molecules produced by the process comprising linking in an operative way a nucleic acid molecule comprising a sequence that hybridizes under stringent hybridization conditions to a sequence set forth in SEQ ID NO:14 and 25, for example, and a sequence controlling the expression of the nucleic acid.

Disclosed are nucleic acid molecules produced by the process comprising linking in an operative way a nucleic acid molecule comprising a sequence encoding a peptide set forth in SEQ ID NO:15 and 26, for example, and a sequence controlling an expression of the nucleic acid molecule.

Disclosed are nucleic acid molecules produced by the process comprising linking in an operative way a nucleic acid molecule comprising a sequence encoding a peptide having 80% identity to a peptide set forth in SEQ ID NO:15 and 26, for example, and a sequence controlling an expression of the nucleic acid molecule.

Disclosed are nucleic acids produced by the process comprising linking in an operative way a nucleic acid molecule comprising a sequence encoding a peptide having 80% identity to a peptide set forth in SEQ ID NO:15 and 26, for example, wherein any change from the SEQ ID NO:15 and 26, for example, are conservative changes and a sequence controlling an expression of the nucleic acid molecule.

Disclosed are cells produced by the process of transforming the cell with any of the disclosed nucleic acids. Disclosed are cells produced by the process of transforming the cell with any of the non-naturally occurring disclosed nucleic acids.

Disclosed are any of the disclosed peptides produced by the process of expressing any of the disclosed nucleic acids. Disclosed are any of the non-naturally occurring disclosed peptides produced by the process of expressing any of the disclosed nucleic acids. Disclosed are any of the disclosed peptides produced by the process of expressing any of the non-naturally disclosed nucleic acids.

Disclosed are animals produced by the process of transfecting a cell within the animal with any of the nucleic acid molecules disclosed herein. Disclosed are animals produced by the process of transfecting a cell within the animal any of the nucleic acid molecules disclosed herein, wherein the animal is a mammal. Also disclosed are animals produced by the process of transfecting a cell within the animal any of the nucleic acid molecules disclosed herein, wherein the mammal is mouse, rat, rabbit, cow, sheep, pig, or primate.

Also disclosed are animals produced by the process of adding to the animal any of the cells disclosed herein.

D. EXAMPLES

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how the compounds, compositions, articles, devices and/or methods claimed herein are made and evaluated, and are intended to be purely exemplary and are not intended to limit the disclosure. Efforts have been made to ensure accuracy with respect to numbers e.g., amounts, temperature, etc.), but some errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, temperature is in ° C. or is at ambient temperature, and pressure is at or near atmospheric.

1. Example 1

Design of Degenerate Oligonucleotides for the Isolation of Desaturase and Elongase from *Thraustochytrid* ONC-T18

Analysis of the fatty acid composition of *Thraustochytrid* ONC-T18 revealed the presence of considerable amount of longer chain PITA such as arachidonic acid (ARA, 20:4n-6), eicosapentaenoic acid (EPA, 20:5n-3), adrenic acid (ADA, 22:4n-6, ω6-docosapentaenoic acid (ω6-DPA, 22:5n-6), ω3-docosapentaenoic acid (ω3-DPA n-3), and docosahexaenoic acid (DHA, 22:6n-3). Thus it was thought that this organism contained an active Δ5-elongase capable of converting ARA to ADA or EPA to ω3-DPA and an active Δ4-desaturase which desaturates ADA to ω6-DPA or ω3-DPA to DHA (FIG. 1). The goal thus was to attempt to isolate these desaturase and elongase genes from *Thraustochytrid* ONC-T18, and eventually to verify the functionality by expression in an alternate host.

To isolate genes encoding functional desaturase enzymes, genomic DNA was extracted from the organism. *Thraustochytrid* ONC-T18 cultures were grown in a growth medium (5 g/l yeast extract, 5 g/l peptone, 40 g/l D(+)-glucose, 1.25 ml/l trace elements, 1.25 ml/l vitamins, 40 g/l sea salt; (trace elements: 5 g/l NaH₂PO₄·H₂O, 3.15 g/l FeCl₃·6H₂O, 4.36 g/l Na₂EDTA·2H₂O, 0.6125 mg/l CuSO₄·5H₂O, 0.05970 g/l Na₂MoO₄·2H₂O, 0.022 g/l ZnSO₄·7H₂O, 0.01 g/l CoCl₂·6H₂O, 0.18 MnCl₂·4H₂O, 13 μg/l H₂SeO₃, 2.7 mg/l NiSO₄·6H₂O, 1.84 mg/l Na₃VO₄, 1.94 mg/l K₂CrO₄), (vitamins: 1 mg/l vitamin B12, 1 mg/l biotin, 0.20 g/l thiamine HCl)) at 26° C. for 16-20 hours with constant agitation. The cells were centrifuged 500 rpm, 5 min at room temperature in Sorvall Super T21 centrifuge with rotor ST-H750 using adapter Sorvall #00436, removed 80% of supernatant, resuspended cells in remaining medium and continued with extraction. Genomic DNA was isolated from cells using Ultraclean Microbial DNA Isolation kit (MO BIO Laboratories, Inc, Solana Beach, Calif.) as per manufacturer's protocol.

The approach taken was to design degenerate oligonucleotides (i.e., primers) that are conserved in known desaturases. These primers could be used in a PCR reaction to identify a fragment containing the conserved regions in the predicted desaturase genes from *Thraustochytrium*. Five sequences were available from *Thraustochytrium* sp., *Thraustochytrium aureum*, *Thraustochytrium* sp. ATCC 34304, *Thraustochytrium* sp. ATCC 21685, and *Thraustochytrium* sp. FJN-10 (EMBO accession number CS020087, Genbank accession number AF391546,

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AF391543, AF489589, DQ133575, respectively). The degenerate primers used were as follows using the Kodon primer designer software: Primer 4desat308 (Forward) (SEQ ID NO:1) 5'-GGRACAGCGASTTTTACAGGG-3', Primer 4desat1369 (Reverse) (SEQ ID NO:2) 5'-GTGCT-CAATCTGGTGGTTKAG-3'.

The same approach was taken to design degenerate oligonucleotides (i.e., primers) that are conserved in known elongases. These primers could be used in a PCR reaction to identify a fragment containing the conserved regions in the predicted elongase genes from *Thraustochytrium* sp. Two sequences were available from *Thraustochytrium* sp, and *Thraustochytrium aureum* (EMBO accession number CS160897 and CS160879, respectively). The degenerate primers used were as follows using Kodon primer designer software: Primer 5elo202F (Forward) (SEQ ID NO:3) 5'-AAGCCYTTTCGAGCTCAAGTYC-3', Primer 5elo768R (Reverse) (SEQ ID NO:4) 5'-GCACGAARAAGTTGC-CGAAG-3'. The degeneracy code for the oligonucleotide sequences was: K=G,T, R=A,G, S=G,C, Y=C,T.

2. Example 2

Isolation of Δ5-Elongase Nucleotide Sequences from *Thraustochytrium* sp. ONC-T18

To isolate the Δ5-elongase gene, PCR amplification was carried out in a 50 μl volume containing: 200 ng *Thraustochytrium* sp. ONC-T18 genomic DNA, 10 μl betaine, 10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, 200 μM each deoxyribonucleotide triphosphate, 1 μM of each primer and 0.2 unit of Taq DNA Polymerase (Sigma, Oakville, Ontario). Thermocycling was carried out at an annealing temperature of 55.7° C., the PCR reaction was resolved on a 0.8% low melt agarose gel, and the band of ~600 bp was gel purified. PCR product was eluted from agarose with water and then purified with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). These DNA fragments were cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue competent cells (Novagen, San Diego, Calif.), and clones were sequenced. One clone was thus isolated that showed sequence homology to previously identified Δ5-elongase. This clone is described as follows: Clone #600-16 (SEQ ID NO:5) was sequenced and the deduced amino acid sequence from 593 bp showed 95% identity with polyunsaturated fatty acid elongase 1 *Thraustochytrium* sp. FJN-10 as the highest scoring match in a BLASTx search.

To isolate the 3'-end, genome walking was carried out using APAGene GOLD Genome Walking kit (BIO S&T, Montreal, Quebec) as per manufacturer's protocol, using genomic DNA purified from *Thraustochytrium* sp. ONC-T18, and oligonucleotides 3'GSPa (SEQ ID NO:6) (5'-CGCTGCGCCCGTACATTACTACCATCCA-3') and 3'GSPb (SEQ ID NO:7) (5'-GTCGTCCAGTCCGTCATGAC-3'). Both oligonucleotides were designed based on the #600-16 fragment of the putative Δ5-elongase. The PCR fragments were resolved on 0.8% low melt agarose gel, and the bands of ~500 to 2000 bp were gel purified. PCR products were eluted from agarose with water and then purified with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). These DNA fragments were cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue

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competent cells (Novagen, San Diego, Calif.), and clones were sequenced. Clone 3'C2-1 (SEQ ID NO:8) contained a 358 bp insert which was identified to contain the 3'-end of the putative Δ5-elongase gene based on sequence homology with known Δ5-elongase and the presence of the 'TGA' stop codon.

To isolate the 5'-end, genome walking was carried out using APAGene GOLD Genome Walking kit (BIO S&T, Montreal, Quebec) as per manufacturer's protocol, using genomic DNA purified from *Thraustochytrium* sp. ONC-T18, and oligonucleotides 5'GSPa (SEQ ID NO:9) (5'-CTCGGCACCCTTCTCCATCGGGTTGCCA-3') and 5'GSPb (SEQ ID NO:10) (5'-GTTGCCAAAGAGCTTG-TAGCCGCCGA-3'). Both oligonucleotides were designed based on the #600-16 fragment of the putative Δ5-elongase. The PCR fragments were resolved on 0.8% low melt agarose gel, and the bands of ~500 to 2000 bp were gel purified. PCR products were dotted from agarose with water and then purified with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). These DNA fragments were cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue competent cells (Novagen, San Diego, Calif.), and clones were sequenced. Clone 5'D2-11 (SEQ ID NO:11) was thus obtained that contained a 519 bp insert that contained the putative 'ATG' start site of the novel Δ5-elongase. The deduced amino acid sequence of this fragment, when aligned with known Δ5-elongase showed 89% identity.

This Δ5-elongase gene was isolated in its entirety by PCR amplification using, the *Thraustochytrium* ONC-T18 genomic DNA as a template, and the following oligonucleotides: ONC-T18elo1 (Forward) (SEQ ID NO:12) 5'-GCT-GATGATGGCCGGGACC-3', ONC-T18elo1099 (Reverse) (SEQ ID NO:13) 5'-GCTCCACTCGAATTCGTAGCG-3'.

PCR amplification was carried out using in a 50 μl volume: 100 ng of the *Thraustochytrium* ONC-T18 genomic DNA, 25 mM TAPS-HCl, pH 9.3, 50 mM KCl, 2 mM MgCl₂, 1 mM β-mercaptoethanol, 1.5 μl DMSO, 200 μM each deoxyribonucleotide triphosphate, 0.5 μM of each primer and 1 unit Phusion high-fidelity DNA polymerase (Finnzymes, Espoo, Finland). Thermocycling conditions were as follows: the template was initially denatured at 98° C. for 30 sec, followed by 30 cycles of [98° C. for 10 sec, 62° C. for 30 sec, 72° C. for 30 sec], and finally an extension cycle at 72° C. for 5 minutes. The PCR product thus obtained was cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue competent cells (Novagen, San Diego, Calif.), and clones were sequenced. The plasmid was purified using UltraClean 6 Minute Mini Plasmid Prep kit (MO BIO Laboratories, Inc, Solana Beach, Calif.). The plasmid thus obtained was digested with BamHI/NotI and cloned into the yeast expression vector pYES2 (Invitrogen, Carlsbad, Calif.) to generate clone pYelo which was then used for expression studies.

The Δ5-elongase full-length gene insert was 1099 bp (SEQ NO:14) in length and, beginning with the first ATG, contained an 831 bp open reading frame encoding 276 amino acids. The amino acid sequence of the full-length gene (SEQ ID NO:15) contained regions of homology to Δ5-elongase from *Thraustochytrium* sp. FJN-10, *Marchantia polymorpha*, *Physcomitrella patens*, and *Mortierella alpina*.

3. Example 3

Isolation of $\Delta 4$ -Desaturase Nucleotide Sequences from *Thraustochytrium* ONC-T18

To isolate the $\Delta 4$ -desaturase gene, PCR amplification was carried out in a 50 μ l volume containing: 100 ng *Thraustochytrium* ONC-T18 genomic DNA, 20 mM Tris-HCl, pH 8.4, 50 mM KCl, 2 mM MgCl₂, 400 μ M each deoxyribonucleotide triphosphate, 2 μ M of each primer and 0.1 unit Tag DNA Polymerase (Invitrogen, Carlsbad, Calif.). Thermocycling was carried out at an annealing temperature of 59.5° C., a portion of the PCR reaction was resolved on a 0.8% agarose gel, and the band of ~1100 bp was purified from the remaining PCR reaction with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). This DNA fragment was cloned into the TOPO TA Cloning kit for Sequencing (Invitrogen, Carlsbad, Calif.) as per manufacturer's specifications. The recombinant plasmid was transformed into TOP10 competent cells (Invitrogen, Carlsbad, Calif.), and clones were sequenced. One clone was thus isolated that showed sequence homology to previously identified $\Delta 4$ -desaturase. This clone is described as follows: Clone #10-3 (SEQ ID NO:16) was sequenced and the deduced amino acid sequence from 967 bp showed 96% identity with delta-4 fatty acid desaturase *Thraustochytrium* sp. ATCC 21685 as the highest scoring match in a BLASTx search.

To isolate the 3'-end, genome walking was carried out using APAGene GOLD Genome Walking kit (BIO S&T Montreal, Quebec) as per manufacturer's protocol, using genomic DNA purified from *Thraustochytrium* ONC-T18, and oligonucleotides 4desat3'a (SEQ ID NO:17) (5'-TTACGCTTCCAAGGACGCGGTC-3') and 4desat3'b (SEQ ID NO:18) (5'-ATGAACAACACGCGCAAGGAGG-3'). Both oligonucleotides were designed based on the #10-3 fragment of the putative $\Delta 4$ -desaturase. The PCR fragments were resolved on 0.8% low melt agarose gel, and the bands of ~500 to 2000 bp were gel purified. PCR products were eluted from agarose with water and then purified with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). These DNA fragments were cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue competent cells (Novagen, San Diego, Calif.), and clones were sequenced. Clone 3'D2-92 (SEQ ID NO:19) contained a 782 bp insert which was identified to contain the 3'-end of the putative $\Delta 4$ -desaturase gene based on sequence homology with known $\Delta 4$ -desaturase and the presence of the 'TGA' stop codon.

To isolate the 5'-end, genome walking was carried out using APAGene GOLD Genome Walking kit (BIO S&T, Montreal, Quebec) as per manufacturer's protocol, using genomic DNA purified from *Thraustochytrium* sp. ONC-T18, and oligonucleotides 4desat5'a (SEQ ID NO:20) (5'-CTGGATACACGTGCCACGAAG-3') and 4desat5'b (SEQ ID NO:21) (5'-CACATCCAGTACAACGAGCTC-CAGAA-3'). Both oligonucleotides were designed based on the #10-3 fragment of the putative $\Delta 4$ -desaturase. The PCR fragments were resolved on 0.8% low melt agarose gel, and the bands of ~500 to 2000 bp were gel purified. PCR products were eluted from agarose with water and then purified with QIAquick PCR Purification kit (Qiagen, Valencia, Calif.). These DNA fragments were cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue

competent cells (Novagen, San Diego, Calif.), and clones were sequenced. Clone 5'-217 (SEQ ID NO:22) was thus obtained that contained a 946 bp insert that contained the putative 'ATG' start site of the novel $\Delta 4$ -desaturase. The deduced amino acid sequence of this fragment, when aligned with known $\Delta 4$ -desaturase showed 96% identity.

This $\Delta 4$ -desaturase gene was isolated in its entirety by PCR amplification using, the *Thraustochytrium* sp. ONC-T18 genomic DNA as a template, and the following oligonucleotides: ONC-T18des380F (Forward) (SEQ ID NO:23) 5'-CGATTGAGAACC GCAAGCTTT-3' ONC-T18DES1687R (Reverse) (SEQ ID NO:24) 5'-GCAGCACTGCTGTGCTCTGGT-3'.

PCR amplification was carried out using in a 50 μ l volume: 300 ng of the *Thraustochytrium* ONC-T18 genomic DNA, 25 mM TAPS-HCl, pH 9.3, 50 mM KCl, 2 mM MgCl₂, 1 mM β -mercaptoethanol, 1.5 μ l DMSO, 200 μ M each deoxyribonucleotide triphosphate, 0.5 μ M of each primer and 1 unit Phusion high-fidelity DNA polymerase (Finnzymes, Espoo, Finland). Thermocycling conditions were as follows: the template was initially denatured at 98° C. for 30 sec, followed by 30 cycles of [98° C. for 10 sec, 61° C. for 30 sec, 72° C. for 30 sec], and finally an extension cycle at 72° C. for 5 minutes. The PCR product thus obtained was cloned into the pT7Blue-3 Perfectly Blunt Cloning kit (Novagen, San Diego, Calif.) as per manufacturer's specifications. The recombinant plasmids were transformed into NovaBlue competent cells (Novagen, San Diego, Calif.), and clones were sequenced, the plasmid was purified using UltraClean 6 Minute Mini Plasmid Prep kit (MO BIO Laboratories, Inc, Solana Beach, Calif.). The plasmid thus obtained was digested with BamHI/NotI and cloned into the yeast expression vector pYES2 (Invitrogen, Carlsbad, Calif.) to generate clone pYDes which was then used for expression studies.

The $\Delta 4$ -desaturase full-length gene insert was 1757 bp (SEQ ID NO:25) in length and, beginning with the first ATG, contained an 1509 bp open reading frame encoding 519 amino acids. The amino acid sequence of the full-length gene (SEQ ID NO:26) contained regions of homology to $\Delta 4$ -desaturase from *Thraustochytrium* sp. ATCC21685, *Thraustochytrium* sp. FJN-10, and *Thraustochytrium aureum*. It also contained the three conserved 'histidine boxes' found in all known membrane-bound desaturases. (Okuley, et al. (1994) The Plant Cell 6:147-158). These were present at amino acid 181-185, 217-222 and 454-458. As with other membrane-bound $\Delta 4$ -desaturases, the third Histidine-box motif (HXXHH(SEQ ID NO:29)) in *Thraustochytrium* ONC-T1 8 $\Delta 4$ -desaturase was found to be QXXHH(SEQ ID NO:30). This sequence also contained a cytochrome b5 domain at the 5'-end. This cytochrome is thought to function as an electron donor in these enzymes.

4. Example 4

Expression of *Thraustochytrium* ONC-T18 $\Delta 4$ -Desaturase and $\Delta 5$ -Elongase Genes in Yeast

Clone pYDes, which consisted of the full length $\Delta 4$ -desaturase cloned into pYES2 (Invitrogen, Carlsbad, Calif.), and clone pYElo, which consisted of the full-length $\Delta 5$ -elongase gene in pYES2, were transformed into competent *Saccharomyces cerevisiae* INVSc1. Yeast transformation was carried out using the S. c. EasyComp Transformation kit (Invitrogen, Carlsbad, Calif.) according to conditions specified by the manufacturer. Transformants were selected for uracil auxotrophy on media lacking uracil

(SC-Ura). To detect the specific desaturase activity of these clones, transformants were grown in the presence of 500 μ M specific fatty acid substrates as listed below: A) Docosapentaenoic acid (22:5n-3) (conversion to docosahexaenoic acid would indicate Δ 4-desaturase activity), B) Docosatetraenoic acid (22:4n-6) (conversion to docosapentaenoic acid (22:5n-6) would indicate Δ 4-desaturase activity), C) Dihomogamma-linolenic acid (20:3n-6) (conversion to arachidonic acid (20:4n-6) would indicate Δ 5-desaturase activity), D) Eicosapentaenoic acid (20:5n-3) (conversion to docosapentaenoic acid (22:5n-3) would indicate Δ 5-elongase activity), E) Arachidonic acid (20:4n-6) (conversion to docosatetraenoic acid (22:4n-6) would indicate Δ 5-elongase activity)

The negative control strain was INVSc1 containing the unaltered pYES2 vector, and these were grown simultaneously. The cultures were vigorously agitated (150 rpm) and grown for 96 hours at 20° C. in the presence of 500 μ M (final concentration) of the various substrates. The cells were pelleted and washed in 100 mM phosphate buffer, pH 7.0, cell pellets were freeze dried. The lipids were then extracted and derivitized to fatty acid methyl esters (FAME) for gas chromatography analysis (GC). Transesterification and extraction was done using 100 mg freeze dried cells, with C19:0 as internal standard, added transesterification reaction mix (methanol:hydrochloric acid:chloroform, 10:1:1) mixed and heated at 90° C. for 2 hours, then allowed to cool at room temperature. FAMES were extracted by adding 1 ml water, and 2 ml hexane:chloroform (4:1), and allow organic and aqueous phases to separate. The organic layer was extracted and treated with 0.5 g of anhydrous sodium sulfate to remove particulates and residual water. The organic solvents were evaporated under a stream of argon. The FAMES were resuspended in iso-octane and analysed by GC-FID. The percent conversion was calculated by dividing the product produced by the sum of (the product produced+ the substrate added) and then multiplying by 100. Table 4 represents the enzyme activity of the genes isolated based on the percent conversion of substrate added.

TABLE 4

Δ 4-desaturase enzyme activity and characterization.					
Vector	Substrate	Product	% conversion		
			average	stdev	
pYES2	C22:5 n-3	C22:6 n-3	0.07	0.14	x = 4
pYDes			14.04	4.01	
pYES2	C22:4 n-6	C22:5 n-6	0.53	0.92	x = 3
pYDes			13.76	1.31	
pYES2	C20:3 n-6	C20:4 n-6	0.42	0.37	x = 3
pYDes			0.87	0.27	

The pYDes clone that contained the Δ 4-desaturase gene from *Thraustochytrium* ONC-T18 converted 14% of the 22:5n-3 substrate to 22:6n-3, as well as 14% of the 22:4n-6 substrate to 22:5n-6. This confirms that the gene encodes a Δ 4-desaturase. There was no background (non-specific conversion of substrate) in this case.

5. Example 5

Manipulation of ONC-T18 Δ 4-Desaturase Activity with Ferric Citrate

Clone pYDes, which consisted of the full length Δ 4-desaturase cloned into pYES2 (Invitrogen, Carlsbad, Calif.), was transformed into competent *Saccharomyces cerevisiae* INVSc1. The negative control strain was INVSc1 containing the unaltered pYES2 vector, and these were grown simultaneously. The cultures were vigorously agitated (150 RPM) and grown for 96 hours at 20° C. in the presence of 500 μ M (final concentration) of DPA and 0.01% ferric citrate. The cells were then treated as described previously to determine the percent conversion of DPA to DHA. The percent conversion, in the presence of ferric citrate was 38.70%, in this case a 2.75 fold increase of Δ 4-desaturase activity.

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          15              20              25

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aac aag ctg aac gag ggc aag ctc ttt atc tcg acc gag gag atg atg 266
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gcg ctc atc gtc ggc tac ctg gcg ttc gtg gtt ctc ggg tct gcc ttc 314
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atg acg gcc ttt gtg agt aag cct ttc gag ctc aag ttc ctg aag ctc 362
Met Thr Ala Phe Val Ser Lys Pro Phe Glu Leu Lys Phe Leu Lys Leu
          65              70              75

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Val His Asn Ile Phe Leu Thr Gly Leu Ser Leu Tyr Met Ala Ser Glu
          80              85              90

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tgc gcg cgc cag gcc tac ctc ggc ggc tac aag ctc ttt ggc aac ccg 458
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          95              100              105

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125                130                135                140

atg atc ctc ggc aag aag tgg aag cag ctc agc ttt ctt cac gtc tac      602
Met Ile Leu Gly Lys Lys Trp Lys Gln Leu Ser Phe Leu His Val Tyr
                145                150                155

cac cac gcg agc atc agc ttc atc tgg ggc att atc gcc cgt ttt gcg      650
His His Ala Ser Ile Ser Phe Ile Trp Gly Ile Ile Ala Arg Phe Ala
                160                165                170

ccc ggt ggc gac gcg tac ttt tcc acc atc ctc aac agc agc gtc cat      698
Pro Gly Gly Asp Ala Tyr Phe Ser Thr Ile Leu Asn Ser Ser Val His
                175                180                185

gtc gtg ctc tac ggc tac tac gcc tcg acc acg ctc ggc tac acc ttc      746
Val Val Leu Tyr Gly Tyr Tyr Ala Ser Thr Thr Leu Gly Tyr Thr Phe
190                195                200

atg cgc ccg ctg cgc ccg tac att act acc atc cag ctc acg cag ttc      794
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270                275

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Gly Tyr Leu Ala Phe Val Val Leu Gly Ser Ala Phe Met Thr Ala Phe
50                55                60

Val Ser Lys Pro Phe Glu Leu Lys Phe Leu Lys Leu Val His Asn Ile
65                70                75                80

Phe Leu Thr Gly Leu Ser Leu Tyr Met Ala Ser Glu Cys Ala Arg Gln
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Ile	Ser	Phe	Ile	Trp	Gly	Ile	Ile	Ala	Arg	Phe	Ala	Pro	Gly	Gly	Asp
				165					170						175
Ala	Tyr	Phe	Ser	Thr	Ile	Leu	Asn	Ser	Ser	Val	His	Val	Val	Leu	Tyr
			180					185						190	
Gly	Tyr	Tyr	Ala	Ser	Thr	Thr	Leu	Gly	Tyr	Thr	Phe	Met	Arg	Pro	Leu
		195					200					205			
Arg	Pro	Tyr	Ile	Thr	Thr	Ile	Gln	Leu	Thr	Gln	Phe	Met	Ala	Met	Val
210						215					220				
Val	Gln	Ser	Val	Tyr	Asp	Tyr	Tyr	Asn	Pro	Cys	Asp	Tyr	Pro	Gln	Pro
225					230					235					240
Leu	Val	Lys	Leu	Leu	Phe	Trp	Tyr	Met	Leu	Thr	Met	Leu	Gly	Leu	Phe
			245					250						255	
Gly	Asn	Phe	Phe	Val	Gln	Gln	Tyr	Leu	Lys	Pro	Lys	Ala	Pro	Lys	Lys
			260					265						270	
Gln	Lys	Thr	Ile												
		275													

<210> SEQ ID NO 16

<211> LENGTH: 276

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; ONCT18_d5elongase peptide

<400> SEQUENCE: 16

Met	Glu	Val	Ala	Gly	Gln	Gln	Trp	Arg	Arg	Phe	Val	Asp	Ala	Val	Asp
1				5					10					15	
Asn	Arg	Ile	Val	Glu	Phe	Met	Glu	His	Glu	Lys	Pro	Asn	Lys	Leu	Asn
			20					25					30		
Glu	Gly	Lys	Leu	Phe	Ile	Ser	Thr	Glu	Glu	Met	Met	Ala	Leu	Ile	Val
		35					40					45			
Gly	Tyr	Leu	Ala	Phe	Val	Val	Leu	Gly	Ser	Ala	Phe	Met	Thr	Ala	Phe
		50				55					60				
Val	Ser	Lys	Pro	Phe	Glu	Leu	Lys	Phe	Leu	Lys	Leu	Val	His	Asn	Ile
65					70					75					80
Phe	Leu	Thr	Gly	Leu	Ser	Leu	Tyr	Met	Ala	Ser	Glu	Cys	Ala	Arg	Gln
				85					90					95	
Ala	Tyr	Leu	Gly	Gly	Tyr	Lys	Leu	Phe	Gly	Asn	Pro	Met	Glu	Lys	Gly
			100					105					110		
Ala	Glu	Ser	His	Ala	Leu	Gly	Met	Ala	Ser	Ile	Ile	Tyr	Val	Phe	Tyr
			115				120						125		
Val	Ser	Lys	Phe	Leu	Glu	Phe	Leu	Asp	Thr	Val	Phe	Met	Ile	Leu	Gly
						135						140			
Lys	Lys	Trp	Lys	Gln	Leu	Ser	Phe	Leu	His	Val	Tyr	His	His	Ala	Ser
145					150					155					160
Ile	Ser	Phe	Ile	Trp	Gly	Ile	Ile	Ala	Arg	Phe	Ala	Pro	Gly	Gly	Asp
				165					170						175
Ala	Tyr	Phe	Ser	Thr	Ile	Leu	Asn	Ser	Ser	Val	His	Val	Val	Leu	Tyr
			180					185						190	
Gly	Tyr	Tyr	Ala	Ser	Thr	Thr	Leu	Gly	Tyr	Thr	Phe	Met	Arg	Pro	Leu

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195				200				205							
Arg	Pro	Tyr	Ile	Thr	Thr	Ile	Gln	Leu	Thr	Gln	Phe	Met	Ala	Met	Val
210						215					220				
Val	Gln	Ser	Val	Tyr	Asp	Tyr	Tyr	Asn	Pro	Cys	Asp	Tyr	Pro	Gln	Pro
225					230					235					240
Leu	Val	Lys	Leu	Leu	Phe	Trp	Tyr	Met	Leu	Thr	Met	Leu	Gly	Leu	Phe
				245					250					255	
Gly	Asn	Phe	Phe	Val	Gln	Gln	Tyr	Leu	Lys	Pro	Lys	Ala	Pro	Lys	Lys
			260						265					270	
Gln	Lys	Thr	Ile												
			275												

<210> SEQ ID NO 17

<211> LENGTH: 967

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; 10-3 4DES clone synthesized from recombinant plasmid

<400> SEQUENCE: 17

```

tacagggatga tgcgcgagcg cgtcgtggcc cggctcaggg agcgcggcaa ggcccgcgcg      60
ggtaggctatg agctctggac taaggcattt ctgctcctcg tgggtttctg gagctcgttg      120
tactggatgt  gcacgctgga ccctcgttc  ggggccatct tggccgccat gtcgctgggc      180
gtctttgccc ccttcgtggg  cacgtgtatc cagcacgacg gcaacctagg cgcttttgcc      240
cagtcgcat  gggcaacaa  ggttgccggg  tggacgcttg  acatgattgg  cgccagtggc      300
atgacgtggg  agttccagca  cgtcctgggc  caccaccctg  acacgaacct  gattgaggag      360
gaaaaatggcc  tgcaaaagg  gagcggcaag  aaaatggaca  ccaagggtggc  cgaccaggag      420
agcgaccctgg  atgtcttctc  cacgtacca  atgatgcgct  tgcaccctgtg  gcacgaaaag      480
cgctgggtacc  accgtttcca  acacatttac  ggcccgttca  tcttcggctt  catgaccatc      540
aacaaggtag  tcacgcagga  cgtcaggtg  gtgctccaca  agcggctctt  ccagattgac      600
gccgagtgcc  ggtacgcgag  tccgatgtac  gtggcgcggt  tctggatcat  gaaggcgcctc      660
acggtgctct  acatgggtggg  cctgccttgc  tattegcagg  gcccggtggca  cggcctcaag      720
ctgttcgcga  tcgcgcactt  ttcgtcggc  gaggtgctgg  cgaccatgtt  cattgtgaat      780
cacatcatcg  aggggtgtct  ttacgcttcc  aaggacgcgg  tcaagggcac  gatggcgcct      840
ccgaagacga  tgcacggcgt  gacgccatg  aacaacacgc  gcaaggaggc  ggaggcgcag      900
gcgtccagtc  tggcgcgtga  gtcaagtcag  tccgctcgac  gactgggccc  ctgtcagtgc      960
cagactt                                           967

```

<210> SEQ ID NO 18

<211> LENGTH: 22

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; primer

<400> SEQUENCE: 18

```

ttacgcttcc aaggacgcgg tc                                           22

```

<210> SEQ ID NO 19

<211> LENGTH: 22

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

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<220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; primer

<400> SEQUENCE: 19

atgaacaaca cgcgcaagga gg 22

<210> SEQ ID NO 20
 <211> LENGTH: 782
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; 3' D2-92 clone synthesized from recombinant plasmid

<400> SEQUENCE: 20

atgaacaaca cgcgcaagga ggcggaggcg caggcgtcca agtccggcgc tgtagtcaag 60
 tcagtcccgc tcgacgactg ggcgctgtc cagtgccaga ctteggtgaa ttggagcgtc 120
 ggctcatggt tctggaacca cttttccggc ggcctcaacc accagatcga gcaccatctg 180
 ttccctgggc tcagccaaga gacgtactac cacattcagg acgtcgttca gtctacctgc 240
 gccgagtacg gagtcccgta ccagcacgaa cctcgtctct ggactgcgta ctggaagatg 300
 ctcgagcacc tccgtcagct tggcaatgag gagaccacag agtcctggca gcacactgcc 360
 tgatgggctg acgagggcgg gtgcgaccaa ttacagttc cttgatcag agaggctcca 420
 caaccagagc acagcagtgc tgcgccggcg tcaatgttcc gtcaattggg tgaacgcct 480
 cgggtgctgc ggtggtgctc ttgtttgatt gcttgcttgc ttgcatgoga gcttggtaca 540
 agaccgagcc acccacagcc cgtccatgca tctggggaac agcttgaagt atgaagtact 600
 acgtaccctg ttaagcttta aaaaaaggtc aagtcttacc atgaatgcta atgacgattc 660
 gctggcgtc atcagaggca accaatctc agcaaaaggc ttgcaaaggc caaaaagaag 720
 tcagcgctag cgccacatgc ataaatcagg gctgtcacc cggaggtaaa cgacgcgtgt 780
 tc 782

<210> SEQ ID NO 21
 <211> LENGTH: 22
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; primer

<400> SEQUENCE: 21

ctggatacac gtgcccacga ag 22

<210> SEQ ID NO 22
 <211> LENGTH: 26
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; primer

<400> SEQUENCE: 22

cacatccagt acaacgagct ccagaa 26

<210> SEQ ID NO 23
 <211> LENGTH: 946
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; 5'-217 clone synthesized from recombinant plasmid

-continued

<400> SEQUENCE: 23

```

cacatccagt acaacgagct ccagaaaccc acgaggagca gaaatgcctt agtccagagc   60
tcatagccac cgcggcgggc cttgcccgcg tccttgagcc gggccacgac gcgctcgcgc   120
atcactttgt aaaagtcgct gttccaogtg tagtaogagg ccgacgggag gcctgaaagc   180
gtccgctttt ccttctcatt tgcgccgect tggccatccg gcagcttccc gatccggtag   240
ttgcgagca cgcgctccga gacgccccgc acatggtacg tctcatacaa cacggtggcc   300
tccttgctg cggccagaag gattatgtcg ccgccgggt gcacgctcgc gaacttggtt   360
acatcgtaga cgagcccgtg gatcgcgcac caggcgatc ccggcttgtt gtgcgcgcg   420
acctgctcga acgagatctc ctgctgtag ccgactgca tcctgacccc tgcgggcagt   480
cctgcctact tcgatcaaat tccttcgct acggtcgcgc gctccagcgt cagttcccc   540
tgctcagtga aaggtccaa agcttgoggt tctcaatgc acccagcgcg tccagcctgg   600
gttttgcgcg ctccggcaag atttactgcc aaggatgacg tcatgctttt cgttctgctg   660
tccaagtagt ggcccgatcc ccggcccaag cgcccagctt tgcttgcgag cgaagctccg   720
cgcccgtaca cacgcgctcc tggctgctag cgccatcgcg tggcggggca tcgaccgcat   780
gacgttgcg tcatcatga agcgtgctg acgtacgtac ctaatggtct acctcgacgg   840
cgatgttcgg gctttgcagc aggatgctca ccccgagggt aaacgacgcg tgttcatctg   900
aattcgtcga caagcttctc gagcctagc tagctctaga ccacac   946

```

<210> SEQ ID NO 24

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; ONC-T184des380F primer

<400> SEQUENCE: 24

```

cgattgagaa ccgcaagctt t   21

```

<210> SEQ ID NO 25

<211> LENGTH: 21

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; ONC-T184des1687R primer

<400> SEQUENCE: 25

```

gcagcactgc tgtgctctgg t   21

```

<210> SEQ ID NO 26

<211> LENGTH: 1758

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: synthetic construct; complete Phusion PCR
T18_g4desat clone synthesized from recombinant plasmid

<220> FEATURE:

<221> NAME/KEY: CDS

<222> LOCATION: (119)..(1678)

<400> SEQUENCE: 26

```

cgattgagaa ccgcaagctt tggagccttt cactgagcag ggggaactga cgctggagcg   60
cgcgaccgta ggcgaaggaa tttgatcgaa gtaggcagga ctgctgcag gggtaaac   118
atg aca gtc ggc tac gac gag gag atc tcg ttc gag cag gtc cgc gcg   166
Met Thr Val Gly Tyr Asp Glu Glu Ile Ser Phe Glu Gln Val Arg Ala
1           5           10           15

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cac aac aag ccg gat gac gcc tgg tgc gcg atc cac ggg ctc gtg tac His Asn Lys Pro Asp Asp Ala Trp Cys Ala Ile His Gly Leu Val Tyr 20 25 30	214
gat gta acc aag ttc gcg agc gtg cac ccg ggc ggc gac ata atc ctt Asp Val Thr Lys Phe Ala Ser Val His Pro Gly Gly Asp Ile Ile Leu 35 40 45	262
ctg gcc gca ggc aag gag gcc acc gtg ttg tat gag acg tac cat gtg Leu Ala Ala Gly Lys Glu Ala Thr Val Leu Tyr Glu Thr Tyr His Val 50 55 60	310
cgg ggc gtc tcg gac gcg gtg ctg cgc aag tac cgg atc ggg aag ctg Arg Gly Val Ser Asp Ala Val Leu Arg Lys Tyr Arg Ile Gly Lys Leu 65 70 75 80	358
ccg gat ggc caa ggc ggc gca aat gag aag gaa aag cgg acg ctt tca Pro Asp Gly Gln Gly Ala Asn Glu Lys Glu Lys Arg Thr Leu Ser 85 90 95	406
ggc ctc tcg tcg gcc tcg tac tac acg tgg aac agc gac ttt tac aaa Gly Leu Ser Ser Ala Ser Tyr Tyr Thr Trp Asn Ser Asp Phe Tyr Lys 100 105 110	454
gtg atg cgc gag cgc gtc gtg gcc cgg ctc agg gag cgc ggc aag gcc Val Met Arg Glu Arg Val Val Ala Arg Leu Arg Glu Arg Gly Lys Ala 115 120 125	502
cgc cgc ggt ggc tat gag ctc tgg act aag gca ttt ctg ctc ctc gtg Arg Arg Gly Gly Tyr Glu Leu Trp Thr Lys Ala Phe Leu Leu Leu Val 130 135 140	550
ggt ttc tgg agc tcg ttg tac tgg atg tgc acg ctg gac ccc tcg ttc Gly Phe Trp Ser Ser Leu Tyr Trp Met Cys Thr Leu Asp Pro Ser Phe 145 150 155 160	598
ggg gcc atc ttg gcc gcc atg tcg ctg ggc gtc ttt gcc gcc ttt gtg Gly Ala Ile Leu Ala Ala Met Ser Leu Gly Val Phe Ala Ala Phe Val 165 170 175	646
ggc acg tgt atc cag cac gac ggc aac cat ggc gct ttt gcc cag tcg Gly Thr Cys Ile Gln His Asp Gly Asn His Gly Ala Phe Ala Gln Ser 180 185 190	694
cga tgg gtc aac aag gtt gcc ggg tgg acg ctt gac atg att ggc gcc Arg Trp Val Asn Lys Val Ala Gly Trp Thr Leu Asp Met Ile Gly Ala 195 200 205	742
agt ggc atg acg tgg gag ttc cag cac gtc ctg ggc cac cac ccg tac Ser Gly Met Thr Trp Glu Phe Gln His Val Leu Gly His His Pro Tyr 210 215 220	790
acg aac ctg att gag gag gaa aat ggc ctg caa aag gtg agc ggc aag Thr Asn Leu Ile Glu Glu Glu Asn Gly Leu Gln Lys Val Ser Gly Lys 225 230 235 240	838
aaa atg gac acc aag gtg gcc gac cag gag agc gac ccg gat gtc ttc Lys Met Asp Thr Lys Val Ala Asp Gln Glu Ser Asp Pro Asp Val Phe 245 250 255	886
tcc acg tac cca atg atg cgc ttg cac ccg tgg cac gaa aag cgc tgg Ser Thr Tyr Pro Met Met Arg Leu His Pro Trp His Glu Lys Arg Trp 260 265 270	934
tac cac cgt ttc caa cac att tac ggc ccg ttc atc ttc ggc ttc atg Tyr His Arg Phe Gln His Ile Tyr Gly Pro Phe Ile Phe Gly Phe Met 275 280 285	982
acc atc aac aag gta gtc acg cag gac gtc gag gtg gtg ctc cac aag Thr Ile Asn Lys Val Val Thr Gln Asp Val Glu Val Val Leu His Lys 290 295 300	1030
cgg ctc ttc cag att gac gcc gag tgc cgg tac gcg agt ccg atg tac Arg Leu Phe Gln Ile Asp Ala Glu Cys Arg Tyr Ala Ser Pro Met Tyr 305 310 315 320	1078
gtg gcg cgg ttc tgg atc atg aag gcg ctc acg gtg ctc tac atg gtg Val Ala Arg Phe Trp Ile Met Lys Ala Leu Thr Val Leu Tyr Met Val 1126	1126

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325	330	335	
ggc ctg cct tgc tat tgc cag ggc ccg tgg cac ggc ctc aag ctg ttc Gly Leu Pro Cys Tyr Ser Gln Gly Pro Trp His Gly Leu Lys Leu Phe 340 345 350			1174
gcg atc gcg cac ttt tgc tgc ggc gag gtg ctg gcg acc atg ttc att Ala Ile Ala His Phe Ser Cys Gly Glu Val Leu Ala Thr Met Phe Ile 355 360 365			1222
gtg aat cac atc atc gag ggt gtc tct tac gct tcc aag gac gcg gtc Val Asn His Ile Ile Glu Gly Val Ser Tyr Ala Ser Lys Asp Ala Val 370 375 380			1270
aag ggc acg atg gcg cct ccg aag acg atg cac ggc gtg acg ccc atg Lys Gly Thr Met Ala Pro Pro Lys Thr Met His Gly Val Thr Pro Met 385 390 395 400			1318
aac aac acg cgc aag gag gcg gag gcg cag gcg tcc aag tct ggc gct Asn Asn Thr Arg Lys Glu Ala Glu Ala Gln Ala Ser Lys Ser Gly Ala 405 410 415			1366
gta gtc aag tca gtc ccg ctc gac gac tgg gcc gct gtc cag tgc cag Val Val Lys Ser Val Pro Leu Asp Asp Trp Ala Ala Val Gln Cys Gln 420 425 430			1414
act tcg gtg aat tgg agc gtc ggc tca tgg ttc tgg aac cac ttt tcc Thr Ser Val Asn Trp Ser Val Gly Ser Trp Phe Trp Asn His Phe Ser 435 440 445			1462
ggc ggc ctc aac cac cag atc gag cac cat ctg ttc cct ggg ctc agc Gly Gly Leu Asn His Gln Ile Glu His His Leu Phe Pro Gly Leu Ser 450 455 460			1510
cac gag acg tac tac cac att cag gac gtc gtt cag tct acc tgc gcc His Glu Thr Tyr Tyr His Ile Gln Asp Val Val Gln Ser Thr Cys Ala 465 470 475 480			1558
gag tac gga gtc ccg tac cag cac gaa ccc tcg ctc tgg act gcg tac Glu Tyr Gly Val Pro Tyr Gln His Glu Pro Ser Leu Trp Thr Ala Tyr 485 490 495			1606
tgg aag atg ctc gag cac ctc cgt cag ctt ggc aat gag gag acc cac Trp Lys Met Leu Glu His Leu Arg Gln Leu Gly Asn Glu Glu Thr His 500 505 510			1654
gag tcc tgg cag cac act gcc tga tgggctgaca agggcgggtg cgaccaattt Glu Ser Trp Gln His Thr Ala 515			1708
acagttcctt gatcacgaga ggctccacaa ccagagcaca gcagtgtgc			1758

<210> SEQ ID NO 27

<211> LENGTH: 519

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic Construct

<400> SEQUENCE: 27

Met Thr Val Gly Tyr Asp Glu Glu Ile Ser Phe Glu Gln Val Arg Ala 1 5 10 15
His Asn Lys Pro Asp Asp Ala Trp Cys Ala Ile His Gly Leu Val Tyr 20 25 30
Asp Val Thr Lys Phe Ala Ser Val His Pro Gly Gly Asp Ile Ile Leu 35 40 45
Leu Ala Ala Gly Lys Glu Ala Thr Val Leu Tyr Glu Thr Tyr His Val 50 55 60
Arg Gly Val Ser Asp Ala Val Leu Arg Lys Tyr Arg Ile Gly Lys Leu 65 70 75 80
Pro Asp Gly Gln Gly Gly Ala Asn Glu Lys Glu Lys Arg Thr Leu Ser 85 90 95

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Gly Leu Ser Ser Ala Ser Tyr Tyr Thr Trp Asn Ser Asp Phe Tyr Lys
 100 105 110

Val Met Arg Glu Arg Val Val Ala Arg Leu Arg Glu Arg Gly Lys Ala
 115 120 125

Arg Arg Gly Gly Tyr Glu Leu Trp Thr Lys Ala Phe Leu Leu Leu Val
 130 135 140

Gly Phe Trp Ser Ser Leu Tyr Trp Met Cys Thr Leu Asp Pro Ser Phe
 145 150 155 160

Gly Ala Ile Leu Ala Ala Met Ser Leu Gly Val Phe Ala Ala Phe Val
 165 170 175

Gly Thr Cys Ile Gln His Asp Gly Asn His Gly Ala Phe Ala Gln Ser
 180 185 190

Arg Trp Val Asn Lys Val Ala Gly Trp Thr Leu Asp Met Ile Gly Ala
 195 200 205

Ser Gly Met Thr Trp Glu Phe Gln His Val Leu Gly His His Pro Tyr
 210 215 220

Thr Asn Leu Ile Glu Glu Glu Asn Gly Leu Gln Lys Val Ser Gly Lys
 225 230 235 240

Lys Met Asp Thr Lys Val Ala Asp Gln Glu Ser Asp Pro Asp Val Phe
 245 250 255

Ser Thr Tyr Pro Met Met Arg Leu His Pro Trp His Glu Lys Arg Trp
 260 265 270

Tyr His Arg Phe Gln His Ile Tyr Gly Pro Phe Ile Phe Gly Phe Met
 275 280 285

Thr Ile Asn Lys Val Val Thr Gln Asp Val Glu Val Val Leu His Lys
 290 295 300

Arg Leu Phe Gln Ile Asp Ala Glu Cys Arg Tyr Ala Ser Pro Met Tyr
 305 310 315 320

Val Ala Arg Phe Trp Ile Met Lys Ala Leu Thr Val Leu Tyr Met Val
 325 330 335

Gly Leu Pro Cys Tyr Ser Gln Gly Pro Trp His Gly Leu Lys Leu Phe
 340 345 350

Ala Ile Ala His Phe Ser Cys Gly Glu Val Leu Ala Thr Met Phe Ile
 355 360 365

Val Asn His Ile Ile Glu Gly Val Ser Tyr Ala Ser Lys Asp Ala Val
 370 375 380

Lys Gly Thr Met Ala Pro Pro Lys Thr Met His Gly Val Thr Pro Met
 385 390 395 400

Asn Asn Thr Arg Lys Glu Ala Glu Ala Gln Ala Ser Lys Ser Gly Ala
 405 410 415

Val Val Lys Ser Val Pro Leu Asp Asp Trp Ala Ala Val Gln Cys Gln
 420 425 430

Thr Ser Val Asn Trp Ser Val Gly Ser Trp Phe Trp Asn His Phe Ser
 435 440 445

Gly Gly Leu Asn His Gln Ile Glu His His Leu Phe Pro Gly Leu Ser
 450 455 460

His Glu Thr Tyr Tyr His Ile Gln Asp Val Val Gln Ser Thr Cys Ala
 465 470 475 480

Glu Tyr Gly Val Pro Tyr Gln His Glu Pro Ser Leu Trp Thr Ala Tyr
 485 490 495

Trp Lys Met Leu Glu His Leu Arg Gln Leu Gly Asn Glu Glu Thr His
 500 505 510

-continued

Glu Ser Trp Gln His Thr Ala
515

<210> SEQ ID NO 28
<211> LENGTH: 519
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: synthetic construct; T18_d4desat peptide

<400> SEQUENCE: 28

Met Thr Val Gly Tyr Asp Glu Glu Ile Ser Phe Glu Gln Val Arg Ala
1 5 10 15
His Asn Lys Pro Asp Asp Ala Trp Cys Ala Ile His Gly Leu Val Tyr
20 25 30
Asp Val Thr Lys Phe Ala Ser Val His Pro Gly Gly Asp Ile Ile Leu
35 40 45
Leu Ala Ala Gly Lys Glu Ala Thr Val Leu Tyr Glu Thr Tyr His Val
50 55 60
Arg Gly Val Ser Asp Ala Val Leu Arg Lys Tyr Arg Ile Gly Lys Leu
65 70 75 80
Pro Asp Gly Gln Gly Gly Ala Asn Glu Lys Glu Lys Arg Thr Leu Ser
85 90 95
Gly Leu Ser Ser Ala Ser Tyr Tyr Thr Trp Asn Ser Asp Phe Tyr Lys
100 105 110
Val Met Arg Glu Arg Val Val Ala Arg Leu Arg Glu Arg Gly Lys Ala
115 120 125
Arg Arg Gly Gly Tyr Glu Leu Trp Thr Lys Ala Phe Leu Leu Leu Val
130 135 140
Gly Phe Trp Ser Ser Leu Tyr Trp Met Cys Thr Leu Asp Pro Ser Phe
145 150 155 160
Gly Ala Ile Leu Ala Ala Met Ser Leu Gly Val Phe Ala Ala Phe Val
165 170 175
Gly Thr Cys Ile Gln His Asp Gly Asn His Gly Ala Phe Ala Gln Ser
180 185 190
Arg Trp Val Asn Lys Val Ala Gly Trp Thr Leu Asp Met Ile Gly Ala
195 200 205
Ser Gly Met Thr Trp Glu Phe Gln His Val Leu Gly His His Pro Tyr
210 215 220
Thr Asn Leu Ile Glu Glu Glu Asn Gly Leu Gln Lys Val Ser Gly Lys
225 230 235 240
Lys Met Asp Thr Lys Val Ala Asp Gln Glu Ser Asp Pro Asp Val Phe
245 250 255
Ser Thr Tyr Pro Met Met Arg Leu His Pro Trp His Glu Lys Arg Trp
260 265 270
Tyr His Arg Phe Gln His Ile Tyr Gly Pro Phe Ile Phe Gly Phe Met
275 280 285
Thr Ile Asn Lys Val Val Thr Gln Asp Val Glu Val Val Leu His Lys
290 295 300
Arg Leu Phe Gln Ile Asp Ala Glu Cys Arg Tyr Ala Ser Pro Met Tyr
305 310 315 320
Val Ala Arg Phe Trp Ile Met Lys Ala Leu Thr Val Leu Tyr Met Val
325 330 335
Gly Leu Pro Cys Tyr Ser Gln Gly Pro Trp His Gly Leu Lys Leu Phe
340 345 350

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Ala Ile Ala His Phe Ser Cys Gly Glu Val Leu Ala Thr Met Phe Ile
 355 360 365

Val Asn His Ile Ile Glu Gly Val Ser Tyr Ala Ser Lys Asp Ala Val
 370 375 380

Lys Gly Thr Met Ala Pro Pro Lys Thr Met His Gly Val Thr Pro Met
 385 390 395 400

Asn Asn Thr Arg Lys Glu Ala Glu Ala Gln Ala Ser Lys Ser Gly Ala
 405 410 415

Val Val Lys Ser Val Pro Leu Asp Asp Trp Ala Ala Val Gln Cys Gln
 420 425 430

Thr Ser Val Asn Trp Ser Val Gly Ser Trp Phe Trp Asn His Phe Ser
 435 440 445

Gly Gly Leu Asn His Gln Ile Glu His His Leu Phe Pro Gly Leu Ser
 450 455 460

His Glu Thr Tyr Tyr His Ile Gln Asp Val Val Gln Ser Thr Cys Ala
 465 470 475 480

Glu Tyr Gly Val Pro Tyr Gln His Glu Pro Ser Leu Trp Thr Ala Tyr
 485 490 495

Trp Lys Met Leu Glu His Leu Arg Gln Leu Gly Asn Glu Glu Thr His
 500 505 510

Glu Ser Trp Gln His Thr Ala
 515

<210> SEQ ID NO 29
 <211> LENGTH: 5
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: synthetic construct; histidine_box1 peptide
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <222> LOCATION: (2)..(2)
 <223> OTHER INFORMATION: X can be any amino acid
 <220> FEATURE:
 <221> NAME/KEY: MISC_FEATURE
 <222> LOCATION: (3)..(3)
 <223> OTHER INFORMATION: X can be any amino acid

<400> SEQUENCE: 29

His Xaa Xaa His His
 1 5

<210> SEQ ID NO 30
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 <213> ORGANISM: Artificial Sequence
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 <223> OTHER INFORMATION: X can be any amino acid
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 <223> OTHER INFORMATION: X can be any amino acid

<400> SEQUENCE: 30

Gln Xaa Xaa His His
 1 5

What is claimed is:

1. A vector comprising a selectable marker and a nucleic acid sequence, wherein the nucleic acid encodes a D5 elongase having at least 97% identity to SEQ ID NO:15 and wherein the elongase activity is retained. 5
2. The vector of claim 1, wherein the D5 elongase comprises conservative changes.
3. The vector of claim 1, wherein the isolated nucleic acid comprises SEQ ID NO:14.
4. An isolated cell, wherein the cell comprises the vector 10 of claim 1.
5. The cell of claim 4, wherein the cell is a eukaryote, a prokaryote, a *Thraustochytrid*, a yeast, or an *Escherichia coli*.
6. The cell of claim 4, wherein the elongase is in the 15 presence of an elongase substrate.
7. The cell of claim 6, wherein the substrate has a concentration of at least 100 μM , 200 μM , 300 μM , 400 μM , 500 μM , 600 μM , 700 μM , 800 μM , 900 μM , or 1000 μM .
8. The cell of claim 4, wherein the cell is a plant or 20 non-human animal cell.

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